Dysbiotic gut microbiota causes transmissible Crohn’s disease-like ileitis independent of failure in antimicrobial defence

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ABSTRACT

Objectives Dysbiosis of the intestinal microbiota is associated with Crohn’s disease (CD). Functional evidence for a causal role of bacteria in the development of chronic small intestinal inflammation is lacking. Similar to human pathology, TNFdeltaARE mice develop a tumour necrosis factor (TNF)-driven CD-like transmural inflammation with predominant ileal involvement.

Design Heterozygous TNFdeltaARE mice and wildtype (WT) littermates were housed under conventional (CONV), specific pathogen-free (SPF) and germ-free (GF) conditions. Microbial communities were analysed by high-throughput 16S ribosomal RNA gene sequencing. Metaproteomes were measured using LC-MS. Temporal and spatial resolution of disease development was followed after antibiotic treatment and transfer of microbial communities into GF mice. Granulocyte infiltration and Paneth cell function was assessed by immunofluorescence and gene expression analysis.

Results GF-TNFdeltaARE mice were free of inflammation in the gut and antibiotic treatment of CONV-TNFdeltaARE mice attenuated ileitis but not colitis, demonstrating that disease severity and location are microbiota-dependent. SPF-TNFdeltaARE mice developed distinct ileitis-phenotypes associated with gradual loss of antimicrobial defence. 16S analysis and metaproteomics revealed specific compositional and functional alterations of bacterial communities in inflamed mice. Transplantation of disease-associated but not healthy microbiota transmitted CD-like ileitis to GF-TNFdeltaARE recipients and triggered loss of lysozyme and cryptdin-2 expression. Monoassociation of GF-TNFdeltaARE mice with the human CD-related Escherichia coli LF82 did not induce ileitis.

Conclusions We provide clear experimental evidence for the causal role of gut bacterial dysbiosis in the development of chronic ileal inflammation with subsequent failure of Paneth cell function.

INTRODUCTION

Crohn’s disease (CD) and ulcerative colitis (UC) are the two main forms of inflammatory bowel disease (IBD). While inflammation in patients with CD mainly occurs in the terminal ileum and is found in the proximal colon only occasionally, inflammation in UC is confined to the colon. Despite unknown aetiologies, chronic activation of the mucosal immune system is triggered by gut commensals.

Genome-wide association studies identified links between disease and genes involved in microbial signalling, underlining the role of gut microbes in IBD pathogenesis. Genetic predispositions and inflammation of the host were shown to induce altered composition and metabolic activity of microbial communities, defined as dysbiosis, and, as
expected, patients with IBD are characterised by dysbiotic changes in the gut microbial ecosystem.1,2 Changes in bacterial communities were shown to be shaped by Paneth cell (PC)-derived antimicrobial peptides.12 PCs are located in the crypt base of the small intestine in close proximity to the epithelial stem cell compartment and loss of PC function is associated with ileal phenotypes of CD.12–14 In IBD treatment, the clinical benefit of antibiotics or faecal-stream diversion supports the pathological role of bacteria.12,15 Faecal microbiota transplantation (FMT) is effective in restoring homeostasis in Clostridium difficile infections.16–19 Consequently, there is rising interest for a therapeutical implementation of FMT in UC and CD. However, evidence from controlled clinical trials is still limited and characteristics of a successful donor microbiota are not yet defined. Moreover, the rationale of introducing new antigen pools in a milieu that has been overreacting to microbial stimuli is questionable. For colitis, the importance of bacteria in disease development has been extensively studied in animal models, for example, by showing reduction of colitis in IL-10−/− mice after antibiotic treatment or the ability of various bacterial strains to induce inflammation in germ-free (GF) colitis models.20–23 Due to the lack of GF models for CD-like ileitis, proof for causality of microbes or dysbiosis in the onset of ileitis is lacking.

In the present study, we assessed the impact of intestinal bacterial communities in a spontaneous model of chronic CD-like ileitis. TNFΔΔARE mice carry a deletion in the tumour necrosis factor (TNF) AU-rich (adenosin-uracil) elements (ARE) leading to Tnf driven transmural inflammation in the distal ileum.24 We previously showed that iron-induced modulation of the microbiota is associated with dramatic changes in disease activity of TNFΔΔARE mice.25 However, mechanistic proof for a causal role of microbe-host interactions in the pathogenesis of CD-like ileitis is missing. Here, we used antibiotics and different hygienic conditions (GF, specific pathogen-free (SPF) or conventional (CONV) housing) to dissect the relationship between microbiota changes and ileitis development in TNFΔΔARE mice. To assess the causal role of dysbiotic microbial communities in ileal inflammation, we performed microbiota transplant experiments with GF-TNFΔΔARE mice and characterised PC functions.

**METHODS**

**Ethics statement**

Animal use was approved by the local institution in charge (Regierung von Oberbayern, approval no. 55.2-1-54-2531-75-10 and 55.2-1-54-2531-99-13). All animals were housed in mouse facilities at the Technische Universität München (School of Life Sciences Weihenstephan).

**Housing**

TNFΔΔARE mice were provided by George Kollias (Alexander Fleming BSRC, Greece), bred in our CONV facility and transferred to SPF via embryo transfer. TNFΔΔARE mice were made GF by hysterectomy (Institute for Laboratory Animal Science; Hannover). Sterility was checked by cultivation of faeces in Luria broth (LB) or Wilkins Chalgen agar (WCA) broth (OXOID) and by microscopic observation of Gram-stained faecal smears every 10–14 days and at sampling. A mould-trap was used to indicate the presence of mold. No contaminations were observed during the experiments. Heterozygous TNFΔΔARE and WT littermates (C57BL/6/N) were kept in CONV, SPF or GF conditions (12 h light/dark cycles at 24–26°C) until the age of 18 weeks. Mice were fed a standard diet (autoclaved R/M-H for SPF and CONV, or M-Z V1124-300 for GF-animals, Sniff, Soest, Germany) ad libitum and were sacrificed by CO2.

**Antibiotic treatment**

CONV-TNFΔΔARE and CONV-WT mice received antibiotics (VM: 0.25 g/L vancomycin and 1.0 g/L metronidazole, Sigma-Aldrich and Fluka) from 8 weeks to 12 weeks of age. Antibiotics were prepared fresh twice a week and administered ad libitum via drinking water in light-protected bottles. Mice were sacrificed 0 weeks, 2 weeks, 4 weeks and 6 weeks after cessation of VM therapy.

**Colonisation of GF mice**

Caecal content from SPF mice was collected and immediately suspended (1:10, weight/volume) in filter-sterilised phosphate buffered saline (PBS)/glycerol (20%), snap-frozen and stored (~80°C). Aliquots were centrifuged (300 g/3 min/4°C) to pellet debris. Supernatants were centrifuged (8000 g/10 min/4°C) and pellets were resuspended in equal volumes of PBS. Each mouse was gavaged at 8 weeks of age with 100 μL caecal microbiota-suspensions of one SPF-donor (approximately 1–5x10^6 cells per mouse, as determined by THOMA counting-chamber). Mice were housed in microbiota-specific isolators with mixed genotypes per cage and sacrificed 4 weeks after colonisation. To investigate ileitis development over time, additional mice were colonised as described above, and samples were collected 1 week, 2 weeks and 4 weeks after colonisation. See online supplementary methods for association of mice with Escherichia coli LF-82.

**Cultivation of intestinal bacteria**

See online supplementary methods.

**Histopathology**

H&E stained terminal ileal and proximal colonic tissue sections were scored (blindly) by assessing lamina propria inflammation, crypt hyperplasia, goblet cell depletion and architectural distortion resulting in a score from 0 to 12.26,27 Images were acquired using Digital microscope M8 (Precipoint GmbH).

**Immunofluorescence staining**

See online supplementary methods.

**Gene expression analysis**

RNA of total ileal tissue was isolated according to manufacturer’s instructions (NucleoSpin RNAII kit; Macherey-Nagel GmbH, KG). Complementary DNA was synthesised from 500 ng RNA using random hexamers and moloney murine leukemia virus (M-MLV) reverse transcriptase (RT) (Promega). RT-PCR was performed using the LightCycler 480 Universal Probe Library System (Roche). For primer sequences and respective probes see supplementary methods. Calculations (2−ΔΔCt method) were done normalised to gapdh.

**16S ribosomal RNA gene sequence analysis**

See online supplementary methods.

**Mass spectrometry and proteome analysis**

See online supplementary methods.

**Statistics**

Statistical analyses were performed with R or Sigma Plot 11.0 using analysis of variance (ANOVA) followed by pairwise
comparison testing (Holm-Sidak test). Graphics were created using GraphPad Prism V5.00. Unless otherwise stated, data are presented as mean±SD and p values below 0.05 were considered to be statistically significant. The Benjamini-Hochberg method was used for adjustment after multiple testing. For visualisation of the relationships between bacterial profiles, non-parametrical multiple dimensional scaling plots were computed using the packages vegan and ade4.

RESULTS

Inflammation in TNFdeltaARE mice is microbiota-dependent

To assess the role of the gut microbiota in the development of CD-like inflammation, we generated GF-TNFdeltaARE mice or treated CONV-TNFdeltaARE mice with antibiotics. Under CONV-housing, TNFdeltaARE mice developed tissue pathology in the terminal ileum at the age of 18 weeks (figure 1A, B). In the absence of microorganisms (GF conditions), TNFdeltaARE mice were free of ileal (figure 1A, B) and colonic disease (as online supplementary figure S1A). Although TNFdeltaARE mice are known as model for CD-like ileitis,24 inflammation of the proximal colon was observed in all CONV-mice (see online supplementary figure S1A). There was no correlation between ileitis and colitis severity, as for example several mice with severe ileitis (score >4) developed moderate colitis (score <4).

We previously showed that ileitis develops gradually in CONV-TNFdeltaARE mice and reaches maximum levels at 12 weeks.29 Using 16S ribosomal RNA sequencing, we characterised faecal bacterial diversity and composition at different ages in TNFdeltaARE and WT littermates. In the absence of inflammation (age of 4 weeks), bacterial phylogenetic make-ups in TNFdeltaARE and WT mice overlapped (figure 1C). Shifts in community structure started to be distinct at the age of 8 weeks, when tissue pathology reaches significance (ileitis score of 3.92±0.52; p<0.01). Parallel to the age-related increase in inflammation, microbial communities from WT diverged from TNFdeltaARE and interindividual differences decreased within genotypes. Main significant differences in composition included increased abundance of Bacteroidaceae, Erysipelotrichaceae, Peptostreptococcaceae and Verrucomicrobiaceae in TNFdeltaARE mice (p<0.001; see online supplementary figure S1B). The increase in Bacteroidaceae was mainly due to three operational taxonomic units (OTUs) closely related to Bacteroides acidifaciens and Bacteroides sartorii.

To assess whether changes in microbial communities can antagonise established inflammation, we treated inflamed CONV-TNFdeltaARE mice with antibiotics starting at 8 weeks of age. Ileitis was significantly reduced after VM-treatment and complete relapse of ileitis was achieved 6 weeks after ending the antibiotic treatment (figure 1D). Colitis activity remained unaltered over the whole period (see online supplementary figure S1C). Tnf-expression levels reflected the dynamic changes in ileal disease activity (figure 1E). Numbers of colony-forming units grown under anaerobic conditions (figure 1F) and total cell counts (see online supplementary figure S1D) showed no change after VM treatment, suggesting growth of VM-resistant bacterial species. However, bacterial diversity was drastically reduced in VM-treated mice (figure 1G). Analysis of phyla distribution in TNFdeltaARE mice showed the predominance of Firmicutes and Bacteroidetes in controls, whereas VM treatment led to a reduction of Bacteroidetes and a bloom in Firmicutes (Lactobacillus spp) and Proteobacteria (E. coli) (figure 1H). Interestingly, bacterial community composition showed rapid resilience in overall composition, that is, phyla distribution returned to pre-VM values 2 weeks after cessation of treatment preceding the return of inflammation.

Disease severity in the distal ileum is associated with loss of PC-associated antimicrobial defence

Based on sequencing, culture and Federation of Laboratory Animal Science Associations (FELASA)-recommened analyses, we observed the presence of murine norovirus,30 Helicobacter, Pasteurella, Syphilus, Trichomonas, Candida, Kazachstania and Chlamydiae spp in the CONV-housing facility (see online supplementary table S1). Hence, to study the role of microbial communities in a strictly controlled environment, we transferred CONV-TNFdeltaARE mice by embryo transfer to a specific pathogen-free (SPF) environment. In contrast to age-matched CONV-mice, SPF-TNFdeltaARE mice developed CD-like ileitis but not colitis (figure 2A, B and see online supplementary figure S2A). Interestingly, we observed a gradient of ileitis severity in animals from the SPF facility, including three main categories of TNFdeltaARE mice: (1) 4 mice out of 20 showed no ileitis, hereon referred to as ‘non-responders’ (NRs); (2) 9 mice had an ileitis score >4 and were defined as ‘responders’ (Rs); and (3) 7 mice with a score between 1 and 4 were defined as ‘low responders’ (lowR). Of note, all non-infamed mice were male and no cage effect was observed. Inflammation-related markers, such as cytokine expression, granulocyte infiltration and PC function were measured in the three ileitis phenotypes and in GF-TNFdeltaARE mice. Similar to CONV-TNFdeltaARE mice, Tnf expression levels (figure 2C) and granulocyte-infiltration into the ileal mucosa (figure 2B, D) reflected the gradual increase in disease activity in SPF-TNFdeltaARE mice. Tnf-transcript levels were reduced and Ly6G-positive granulocytes were totally absent in GF-TNFdeltaARE mice.

To assess PC function, lysozyme and α-fucosylated proteins (as detected by Ulex europaeus agglutinin-1 (UEA-1)) were stained in the ileal mucosa of SPF-TNFdeltaARE and WT littermates (figure 3A and see online supplementary figure S2B). Immunofluorescence analysis clearly demonstrated a significant loss of lysozyme but not UEA-1 positive cells in the crypt base of inflamed lowR and R SPF-TNFdeltaARE mice compared with GF controls (figure 3B), while numbers of lysozyme and UEA-1 positive cells in NR TNFdeltaARE mice were comparable to WT. (dotted line). These data suggest a loss of PC activity but not PC number in the presence of inflammation. Besides the loss of lysozyme expression, signals for Cryptdin 2-positive cells were also reduced in severe ileitis (figure 3A, B). Consistent with a loss of PC function in R mice, transcript levels of the antimicrobial peptide Angiogenin 4 was significantly reduced compared with NRs (see online supplementary figure S2C). While Cryptdin 5 (Defensin a5) showed high interindividual variations but the same trend towards reduction under inflammation (p>0.05), Reg γ expression remained unchanged (see online supplementary figure S2C). GF mice showed lower transcript levels of the tested antimicrobial peptides, which accords previous publications.31 32 An increase in the number of Caspase-3-positive cells in the ileal crypts could not be observed (figure S2D).

Ileitis severity is linked to compositional and functional dysbiosis

To investigate the relationship between ileitis severity and the gut microbiota in SPF mice, we performed 16S ribosomal RNA gene sequencing. There was no change in α-diversity between the three ileitis phenotypes (figure 4A). However, β-diversity analysis revealed clear separation of R TNFdeltaARE-mice and their WT littermates (figure 4B). Most interestingly, bacterial community structures of NR TNFdeltaARE-mice clustered together with WT...
Figure 1  Changes in the gut microbial ecosystem are associated with inflammation in TNFdeltaARE mice. (A) Ileitis scores in 18-week-old TNFdeltaARE (ARE) and WT littermates in CONV or GF housing. Ileitis-score groups are colour-coded according to inflammation severity: score=0 (blue); score <4 (grey); score >4 (red). Male mice are displayed as triangles, female mice as circles. (B) Representative H&E-stained sections of the distal ileum. (C) NMDS plot showing shifts of centroids (indicated by arrows) and variance of faecal bacterial profiles (circled areas) of CONV-TNFdeltaARE and WT mice at 4 weeks, 8 weeks and 12 weeks of age. 16S ribosomal RNA gene amplicons of the V3/V4 region (407 bp) in faeces were sequenced on a MiSeq platform. (D) Ileitis scores in TNFdeltaARE mice treated for 4 weeks with vancomycin and metronidazole (VM), starting at 8 weeks of age. Recurrence of ileitis after VM therapy was followed for 6 weeks (n=5–6/group). (E) Ileal expression of Tnf as fold-increase to CONV-WT mice (normalised to Gapdh). (F) Ileal bacterial density of cultivable anaerobes (as log 10 of colony forming units (cfu) per gram ileal content). (G) Intestinal bacterial diversity (Shannon effective counts). (H) Changes in caecal bacterial composition (relative abundance of total sequences) at the phylum level. *p<0.05; **p<0.01; ***p<0.001 (Two-way-ANOVA followed by Holm-Sidak test). CONV, conventional; GF, germ-free; ARE, AU-rich elements; TNF, tumour necrosis factor; NMDS, non-parametrical multiple dimensional scaling.
mice but were separated from R TNFδARE mice. This clearly indicates a link between inflammation and bacterial communities.

Relative abundance of taxonomic groups revealed that changes in bacterial diversity were associated with changes in composition (figure 4C). Ileitis scores negatively correlated with relative abundances of Porphyromonadaceae (r = −0.79; p < 0.001). In contrast, unknown members of the order Clostridiales showed significantly higher proportions in R mice and positively correlated with the degree of histopathology (r = 0.48; p = 0.016).

Additional sequence analysis of faecal samples directly after weaning demonstrated that the transfer of microbiota from mother to offspring is not related to the disease susceptibility later in life and disease-related changes in microbiota are independent of littermate effects. TNFδARE mice of the same litter were found in the three groups of ileitis severity and thus developed the dysbiotic or non-dysbiotic caecal microbiota post weaning (see online supplementary figure S3A).

These differences in composition were underlined by functional changes in microbial communities as determined by LC-MS/MS metaproteome analysis. Partial least squared analysis showed a clear separation of R and NR mice (figure 4D).

Proteins were assigned to functions using clusters of orthologous groups. The subroles translation ribosomal structure and biogenesis (fold change (fc) 103, p = 0.028), carbohydrate transport and metabolism (fc 59.8, p = 0.018) as well as amino acid transport and metabolism (fc 23.4, p = 0.027) showed significant increase in the R group. Proteins that were significantly different between the two groups of mice within the categories metabolism or cellular processes and signalling are shown in figure 4E and see online supplementary figure S3B. Ten proteins were exclusively found in Rs, while only two were specific for NRs.

Ileitis is transmissible via dysbiotic gut microbiota

GF-TNFδARE mice are free of intestinal inflammation, and the occurrence of ileitis in SPF-housing is associated with dysbiosis. We first tested mono-colonisation of GF-TNFδARE mice with the Gram-negative adherent invasive bacterium E coli LF82, a well-known strain associated with CD.33 34 After 4 weeks of colonisation, LF82 did not induce ileitis in TNFδARE mice (p = 0.07), although cell density reached 10.2 ± 0.3 log10 cfu/g in the caecum (see online supplementary figure S4A).
Figure 3  Disease severity is associated with loss of PC function. (A) Paneth cell staining of lysozyme (green; upper panel) combined with UEA-1 (red). A representative crypt is shown in the magnification. Lower panel: Paneth cell staining of cryptin 2 (red). A representative crypt is shown in the magnification. (B) Quantification of lysozyme, UEA-1 and cryptin-2 positive cells per crypt base in ARE-mice compared with WT mice (dotted line). **p<0.01; ***p<0.001; Two-way ANOVA followed by Holm-Sidak test.
To test the causal relationship of a dysbiotic microbiota associated with development of CD-like ileitis in TNF$^{\text{deltaARE}}$ mice, we next transferred caecal microbial communities from SPF-TNF$^{\text{deltaARE}}$ mice into GF-recipients (figure 5A). Therefore, GF-TNF$^{\text{deltaARE}}$ and GF-WT mice were colonised with donor-microbiota from either Rs or NRs. Donor-microbiota is highlighted with halos in figure 4B. TNF$^{\text{deltaARE}}$ mice colonised with dysbiotic microbiota from Rs (+R; red) developed inflammation after 4 weeks, mimicking ileitis severity of corresponding donors (recipient mice and respective donors are shown with the same symbols). WT littermates showed no signs of inflammation, supporting the fact that genetic susceptibility was required for the development of ileitis after transfer. Most remarkably, GF-TNF$^{\text{deltaARE}}$ mice colonised with the microbiota from NRs also showed no signs of ileitis (+NR; blue). GF-mice were colonised by comparable densities of bacteria in both recipient groups (approximately $10^9$ cfu/g) and tissue maturation was confirmed by reduced caecum weight (see online supplementary figure S4B). Mesenteric lymph node weight to body weight ratios were significantly increased in both recipient groups when compared with GF-TNF$^{\text{deltaARE}}$ mice (figure 5B), suggesting the presence of mucosal immune cell activation independent of tissue pathology. Increased Tnf-transcript levels ($p<0.001$) correlated with the induction of tissue pathology (figure 5C).

To verify the hypothesis that transmissible ileitis was due to the transfer of dysbiotic bacterial communities, we analysed caecal content in recipient mice using high-throughput sequencing. α-Diversity analysis showed no differences between genotypes and donor groups (see online supplementary figure S4C). β-Diversity analysis displayed again a clear separation between inflamed and non-inflamed recipients (figure 5D). Differences within the recipient groups could still be assigned to the
The increase in *Clostridiales* spp and decrease in *Porphyromonadaceae* observed in inflamed donor SPF-TNFdeltaARE mice also occurred in recipients, but results did not reach significance (see online supplementary figure S4D). Differential abundance analysis identified two OTUs classified as *Hungatella* sp exclusively present in NRs, with relative abundance of up to 15%. Three other OTUs of the same genus were only present in Rs (figure 5E), indicating species-specific differences between R and NR mice.

**Inflammation precedes loss of PC function**

To better understand the role of PCs in ileitis development, we analysed time series of tissue pathology and PC function in the transfer model of CD-like ileitis. GF-TNFdeltaARE and GF-WT recipients were colonised at the age of 8 weeks with dysbiotic microbiota from R SPF-TNFdeltaARE mice for 1 week, 2 weeks and 4 weeks. As shown in figure 6A, ileitis was absent after 1 week (0.13±0.28), but increased with the duration of colonisation in TNFdeltaARE but not in WT mice. Tnf mRNA expression levels were already significantly increased relative to GF-controls (figure 6C; p<0.001), whereas granulocyte infiltration and mesenteric lymph node weight increased later, parallel to disease severity in TNFdeltaARE mice (figure 6D and see online supplementary figure S5A). The influence of colonisation on caecum-weight and mucus-fucosylation was already visible 1 week post colonisation showing increased staining across the crypt-villus axis (figure 7A; upper panel and see online supplementary figure S5B). Numbers of UEA-1-positive crypt cells remained constant over the entire colonisation period. Most importantly, loss of lysozyme-positive crypt cells of recolonised TNFdeltaARE mice did not preclude the onset of tissue pathology (figure 7B), suggesting that loss of PC function is associated but not causal for the development of CD-like ileitis. Quantification of Cryptdin 2-positive cells per crypt (figure 7A lower panel and 7B right panel) showed no reduction in Cryptdin 2 expression in mice after 4 weeks of colonisation, despite the presence of moderate histopathology and inflammatory activation (increased Tnf expression levels and Ly6G-positive granulocyte infiltration).

**DISCUSSION**

In the present work, we provide for the first time evidence for disease-related causality of bacterial dysbiosis in experimental CD-like ileitis. We showed that GF-TNFdeltaARE mice are protected from intestinal inflammation, and thereby demonstrated the essential role of microbial triggers in a model that shares main clinical features of CD. In several models of colitis, the disease-free status under GF conditions has been shown, for example, in IL-10−/− or T-bet(−/−) × Rag2(−/−) ulcerative colitis (TRUC) mouse model (TRUC) mice and HLA/B27-β2m transgenic rats.23 35 36 However, there are only few IBD-related ileitis models including TNFdeltaARE and SAMP1/YitFc mice.37 GF housing induced attenuated or no disease activity in SAMP1/YitFc mice, but information about the specific contribution of microbial communities in ileitis development is lacking.38 39 Sequential sampling of faecal microbiota demonstrated the gradual influence of inflammation, as illustrated by increasing divergence of bacterial communities over time in TNFdeltaARE versus WT littermates. Several taxa were significantly more abundant in TNFdeltaARE mice, including members of the family *Bacteroidaceae*, which are frequently reported to

**Figure 4** Continued.
increase the risk for the development of intestinal inflammation in humans and mice.29 40–42

Changes in microbial composition induced by antibiotic treatment established a disease course with recurrent inflammation, similar to observations in patients with IBD.43 The clinical efficacy of microbial therapy in CD, including antibiotics, probiotics and FMT is not well established. Meta-analyses show no clear advantage of probiotics in maintenance of remission in patients with CD, but hint towards a beneficial effect of antibiotics.44 45 Interestingly, the development of colitis in TNFdeltaARE mice was only detected in the presence of a variety of mouse-related pathogens in the CONV housing and was not affected by antibiotic treatment, suggesting different mechanisms in the pathogenesis of ileitis and colitis in this model of CD-like inflammation.

Embryo transfer of CONV-TNFdeltaARE mice to a barrier-controlled environment led to the progression of extreme ileal phenotypes, that is, approximately half of SPF-TNFdeltaARE mice developed severe ileitis, whereas 20% remained ileitis-free. Disease activity and inflammatory tissue activation, measured at the level of granulocyte infiltration and Tnf-expression, significantly correlated with the reduction in lysozyme-positive or cryptdin 2-positive PCs and ileal ANG4 expression. PCs play an important role in mucosal defence and were shown to be functionally impaired in IBD, especially in patients with CD.46–49 Genetic risk variants like autophagy-associated ATG16L1-T300A and endoplasmic reticulum stress responses contribute to PC dysfunction and epithelial cell-specific knockout models, that is, Caspase-8−/− or ATG16L1−/− and XBP-1−/− mice, allowed a mechanistic understanding of CD-like disease phenotypes.50–52 Interestingly, these newly generated mouse models share loss of PC functions as an important mechanism of ileal pathogenesis. However, lineage-specific ablation of PCs showed no specific IBD-associated phenotype, suggesting that loss of PC function alone is not sufficient to initialise the onset of disease, but rather facilitates the loss of complex regulatory circuits in the epithelium.53 Interferon-γ inhibits the release of PC-derived antimicrobial peptides,54 supporting our observation that inflammation-associated mediators abrogate PC functions. Therefore, loss of PC functions associated with reduced production of antimicrobial peptides in the small intestine might act as a disease accelerator. We showed that TNF-driven ileitis severity in TNFdeltaARE mice is linked to reduced PC function rather than numbers using combined lysozyme-UEA-1 and cryptdin-2 staining. Time-series analysis of transfer experiments with disease-conditioning microbiota in GF recipients clearly

Figure 5  Ileitis is transmissible via dysbiotic gut microbiota. (A) GF-WT and GF-TNFdeltaARE mice were colonised at the age of 8 weeks with the complex caecal microbiota from SPF responder (+R) or non-responder (+NR) donor-TNFdeltaARE mice (orange halos in figure 4; each n=3). Age-matched and time-matched GF-WT and GF-TNFdeltaARE mice served as controls. Ileitis was scored after 4 weeks of colonisation. SPF-donor mice and respective recipients are shown with identical symbols. (B) Increase in mesenteric lymph node (MLN) weight. (C) Ileal expression (normalised to Gapdh of Tnf as fold-increase to SPF-WT mice. (D) NMDS plot showing clear separation between R (red) and NR (blue) recipients according to respective donor (R1–3 or NR1–3; donor and recipient are displayed using identical symbols). ARE and WT are shown as filled and open symbols, respectively. (E) OTUs associated with R or NR colonisation. Each column represents one recipient ARE mouse. Donors and percentages of total sequences (coloured scale) are indicated below the table. ***p<0.001; Two-way ANOVA followed by Holm-Sidak test. ARE, AU-rich elements; GF, germ-free; NMDS, non-parametrical multiple dimensional scaling.

demonstrated that loss of PC function was subsequent to TNF-driven development of inflammation, suggesting that loss in PC function might contribute to, but seems not to be causal for the divergence of disease-related bacterial communities or development of dysbiosis. SPF-TNF\textsuperscript{\Delta ARE} mice with severe ileitis and loss of antimicrobial peptides were characterised by a taxonomically and functionally dysbiotic gut microbiota. Analysis of intestinal communities showed an ileitis-associated increase in the relative abundance of unknown Clostridiales and reduced abundance of Porphyromonadaceae (order of Bacteroidales). While members of Clostridiales were found in lower abundance in patients with CD, they were increased in TNF\textsuperscript{\Delta ARE} ileitis and TM-IEC-C1gal\textsuperscript{−/−} colitis models, indicating model-specific traits of this very broad taxonomic group.\textsuperscript{55, 56} Consistent with changes in our metaproteome analysis, shotgun metagenomic analysis of microbiota from patients with CD and UC showed functional shifts especially in ileal CD, driven by pathways of carbohydrate transport and mechanism and amino acid biosynthesis.\textsuperscript{57} Metaproteomics revealed that, among others, the abundance of fucose-utilising enzyme was exclusive for the dysbiotic ecosystem in SPF-TNF\textsuperscript{\Delta ARE} mice. Inflammation-driven increase in host-derived fucose was shown to affect metabolic functions of fucosidase-expressing commensal bacteria.\textsuperscript{58} Together with the fact that loss of function in the human Fut2 gene was shown to be associated with variations in the gut microbiota of patients with CD, we might speculate a role for the fucose metabolising enzymes in generating a disease-associated microbial environment in the gut.\textsuperscript{59, 60}

We showed that inflammation is associated with the development of dysbiosis and, most importantly, microbiota transfer experiments confirmed a causal relationship between microbial
dysbiosis and disease initiation in TNFδARE mice. We demonstrated for the first time that a compositionally and functionally diverse microbiota, which originates from donors with the same genetic background but different pathologies, transmits ileal inflammation. Garrett et al demonstrated in TRUC mice that a genotype-specific microbiota transferred colitis to RAG2−/− recipient mice. However, WT cagemates were also inflamed, suggesting a rather pathogenic than dysbiotic trait.61 Interestingly, Powell et al62 identified Helicobacter typhlonius as a key driver of pathogenesis in TRUC mice. In our experiments, inflammation occurred only after transfer of disease-related microbiota to GF-TNFδARE and not WT recipient mice, supporting the necessity of genetic susceptibility for ileitis development, as well as the non-infectious nature of the dysbiotic

**Figure 7** Loss of antimicrobial peptides is secondary to the development of inflammation. (A) Paneth cell staining of lysozyme, UEA-1 and Cryptdin 2, as labelled in the figure (upper panel, 10× magnification; others, 120×). (B) Quantification of lysozyme-positive, UEA-1-positive and cryptdin 2-positive cells per crypt base in AU-rich elements (ARE) mice compared with WT mice (dotted line). *p<0.05; ***p<0.001 (One-way-ANOVA followed by Tukey’s test).
microbial communities. *Vice versa*, the microbiota from NR animals failed to induce CD-like inflammation in GF-TNF$^{\Delta \text{ARE}}$ mice, clearly demonstrating the disease-related specificity of the dysbiotic microbiota.

Again, the overall community structure between recipients of R and NR microbiota were very distinct. OTU-based analysis confirmed that members of the family *Porphyromonadaceae* were increased in NRs, but also that the relative abundance of *Hagatella* spp was significantly different. Of note, phylogeny of the genus *Hagatella* is not yet well established and ambiguous delineation from neighbouring species (e.g., *Clostridium clostridiforme, Clostridium aerotolerans, Clostridium bolteae* and *Clostridium xylanolyticum*) calls for taxonomic amendments.64 65 Hence, different representatives of *Hagatella* were found in higher abundances in R and NR TNF$^{\Delta \text{ARE}}$ mice, which hints at dysbiotic conditions characterised by subtle differences in bacterial profiles down to at least the species level. There is rising evidence that protective or deleterious effects of intestinal bacteria are strain-specific or species-specific, for example, PrtP expression in *Lactobacillus casei*, the presence of polysaccharide A in *Bacteroides fragilis*, or gelatinase E production by colitogenic *Enterococcus faecalis*.66 Some specific pathobionts, for example, *Bilophila wadsworthia* in IL-10$^{-/-}$ mice, have been selected from the commensal microbiota with the capability to transfer colitis into susceptible GF hosts.67 68 However, we failed to induce ileitis in monoassociated TNF$^{\Delta \text{ARE}}$ mice using the CD-related pathobiont *E. coli* LF82.67 68 All our results point towards a community effect of the complex microbiota and loss of aggressive, or gain of protective mechanisms, rather than the selection of aggressive phenotypes as single agents causing the development of CD-like ileitis in TNF$^{\Delta \text{ARE}}$ mice.

In summary, we report that TNF-driven chronic inflammation with CD-like ileal pathology depends on microbial triggers and that inflammation is transmitted via dysbiotic communities of commensal bacteria. Consistent with previously published data, maternally inherited factors seem not relevant to the initial development of dominant dysbiotic bacterial communities and disease susceptibility later in life.69 Understanding the true nature of a dysbiotic and disease-conditioning microbiota seems of essential importance to judge the risk of relapse in patients with IBD after therapeutic intervention or to achieve best possible clinical efficacy in FMT trials. The transfer of IBD-related microbiota in GF-TNF$^{\Delta \text{ARE}}$ mice will be an excellent tool to gain translational insights into the role of complex microbial communities in the progression of CD-like inflammation.

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Data sharing statement We would be happy to make our sequencing data available.

Ethics The animal experiments were performed according to the German guidelines for animal care.

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