

1 Supplementary Methods

2 **Isolation of single cells from GC tissues**

3 Fresh tissues were washed 3 times with Hank's solution containing 1% fetal calf serum before
4 being cut into small pieces. The specimens were then collected in RPMI-1640 medium containing
5 1 mg/ml collagenase IV and 10 mg/ml deoxyribonuclease I and mechanically dissociated using
6 the gentle MACS Dissociator (Miltenyi Biotec). Dissociated cell suspensions were further incubated
7 for 1 h at 37°C under continuous rotation. The cell suspensions were then filtered through a 70 µm
8 cell strainer (BD Labware). Cell viability, as determined by trypan blue exclusion staining, was
9 typically >90%.

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11 **Immunohistochemistry**

12 Paraformaldehyde-fixed and paraffin-embedded samples were cut into 5 µm sections. For
13 immunohistochemical single-staining, the sections were incubated with mouse anti-human CD15,
14 anti-human proliferating cell nuclear antigen (PCNA), or rabbit anti-human CD3 antibodies
15 respectively, and then were stained by horseradish peroxidase (HRP) anti-mouse immunoglobulin
16 G (IgG) or using EnVision G2 System/AP Rabbit/Mouse (Permanent Red) followed by
17 diaminobenzidine. All the sections were finally counterstained with haematoxylin and examined
18 using a microscope (Nikon Eclipse 80i; Nikon).

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20 **Immunofluorescence**

21 Paraformaldehyde-fixed sections of tumour tissues from GC patients were washed in PBS and
22 blocked for 30 min with 20% goat serum in PBS and stained for CD15, and CD3 and/or EpCam,
23 and GM-CSF. Slides were examined with a confocal fluorescence microscope (LSM 510 META,
24 Zeiss).

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26 **Flow cytometry**

27 Flow cytometric analysis was performed according to standard protocols. For intracellular cytokine
28 measurements, the cells were stimulated for 5 h with phorbol myristate acetate (50 ng/ml) plus
29 ionomycin (1 µg/ml) in the presence of GolgiStop. Intracellular cytokine staining was performed

1 after fixation and permeabilization using Perm/Wash solution. The cells were analyzed by
2 multicolour flow cytometry with FACSCanto II (BD Biosciences). Data were analyzed with Flowjo
3 software (TreeStar) or FACSDiva software (BD Biosciences).

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5 **Real-time PCR**

6 RNA of mouse tumours was extracted using PureLink™ FFPE Total RNA Isolation Kit (Invitrogen).
7 The RNA samples were reversed transcribed to cDNA with PrimeScript™ RT reagent Kit
8 (TaKaRa). Primers and probes for granzyme B and perforin were obtained from ThermoFisher
9 (granzyme B, Hs00188051_m1, amplification region exons 4–5; perforin, Hs00169473_ml,
10 amplification region exons 2–3). Real-time PCR was performed on the IQ5 (Bio-Rad). Human
11 GAPDH served as the normalizer. The relative gene expression was expressed as fold change
12 calculated by the $\Delta\Delta C_t$ method.

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14 **ELISA**

15 Human gastric tissues from specimens were collected, homogenized in 1 ml sterile Protein
16 Extraction Reagent, and centrifuged. Tissue supernatants were collected for ELISA.
17 Concentrations of GM-CSF in the tissue supernatants were determined using ELISA kits according
18 to the manufacturer's instructions. Proteins from mouse tumours were extracted using FFPE Total
19 Protein Extraction Kit (Sangon Biotech) according to the manufacturer's instructions. Tumour tissue
20 supernatants were collected for ELISA. Concentrations of perforin and granzyme B in the tumour
21 tissue supernatants were determined using ELISA kits according to the manufacturer's instructions.

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23 **Western blot analysis**

24 Western blot assays were performed on 10%-15% SDS-PAGE gels using equivalent amounts of
25 cell lysate proteins of samples. Five percent skimmed milk or three percent BSA was used for
26 blocking the PDF membranes. Human STAT3 and p-STAT3 were detected with rabbit anti-STAT3
27 and rabbit anti-p-STAT3 antibodies respectively. This was followed by incubation with HRP-
28 conjugated secondary antibodies. Bound proteins were visualized by using SuperSignal® West
29 Dura Extended Duration Substrate kit.

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2 **Microarray experiments**

3 Gene expression profiles of human tumour tissues from GC patients were analyzed with the
4 Affymetrix GeneChip Human Gene 1.0 ST Array (Affymetrix), strictly following the manufacturer's
5 protocol. Microarray experiments were performed at the Genminix Informatics (China) with the
6 microarray service certified by Affymetrix.

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