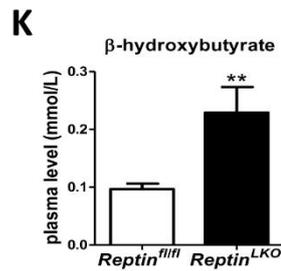
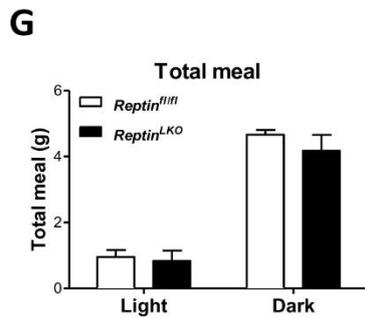
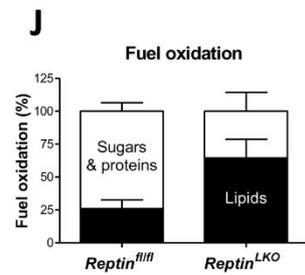
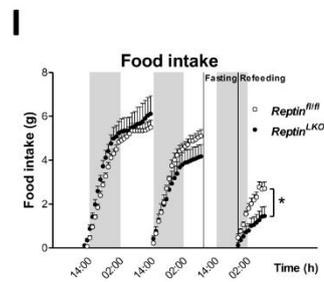
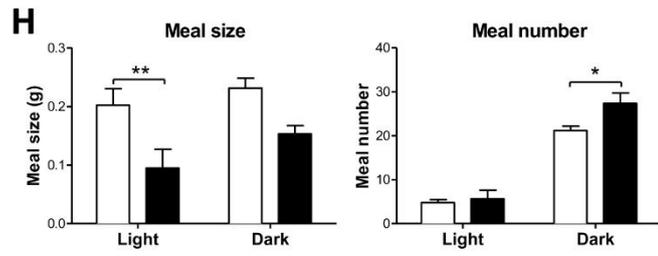
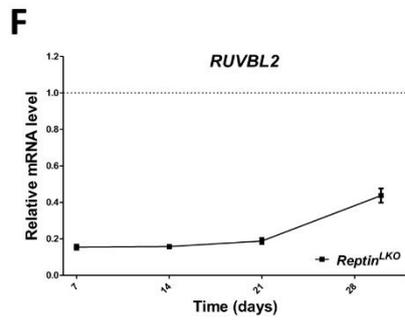
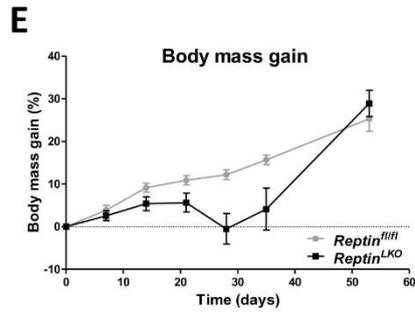
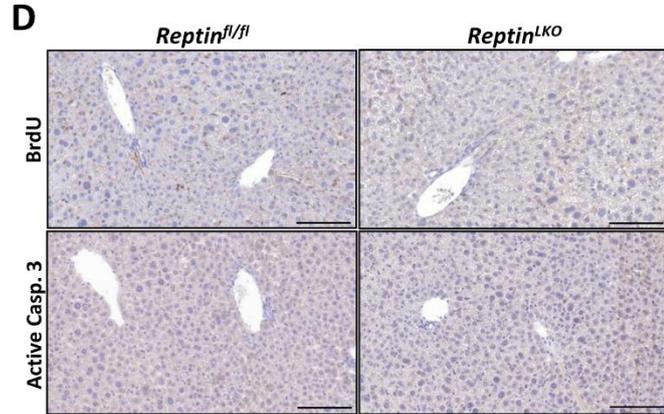
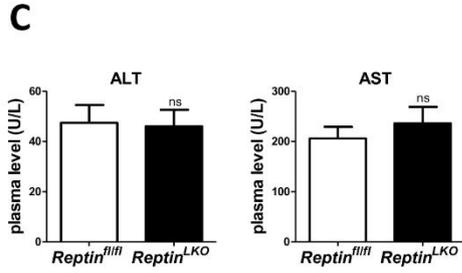
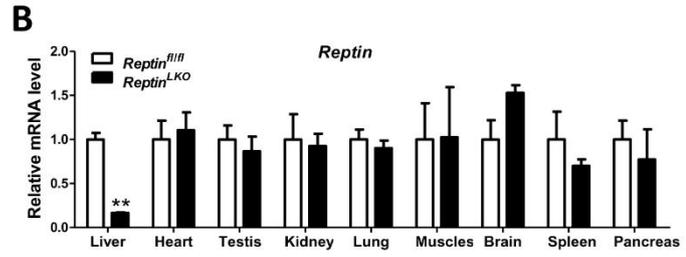
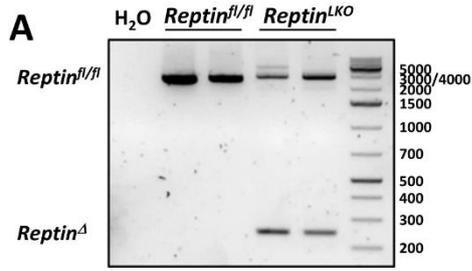


## SUPPLEMENTARY FIGURES

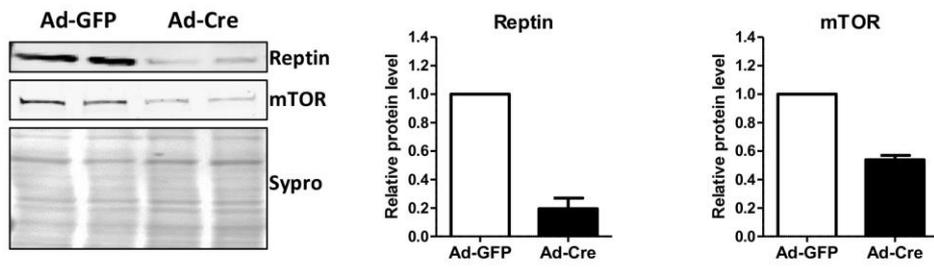
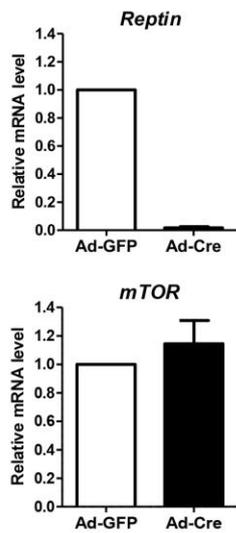
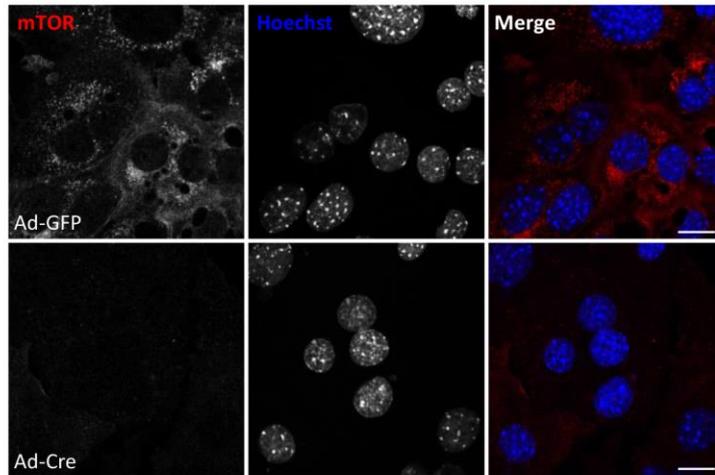
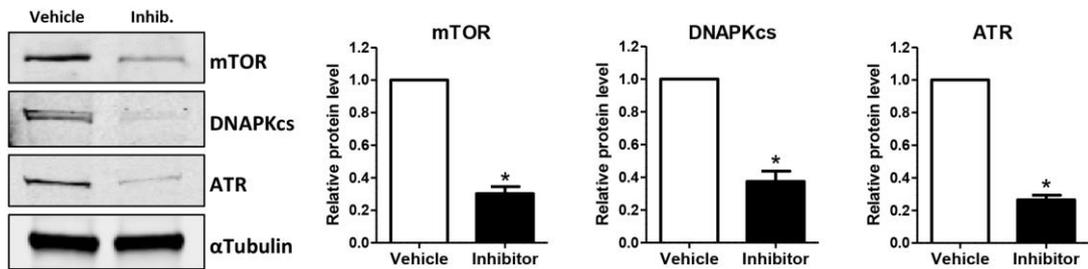
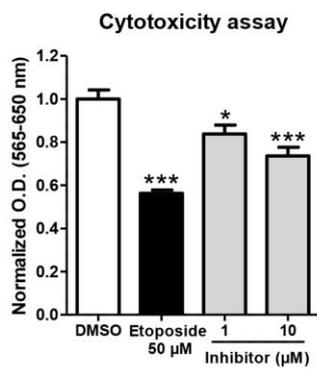
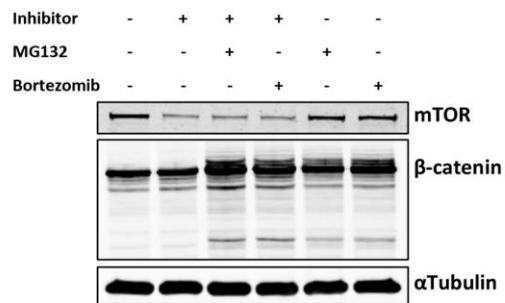
### Sup Figure 1: Characterization of liver-specific Reptin knock-out mice (*Reptin*<sup>LKO</sup>)

**(A)** PCR analysis of *Reptin* gene on genomic DNA extracted from whole livers of *Reptin*<sup>fl/fl</sup> and *Reptin*<sup>LKO</sup> mice at D7 post-tamoxifen injection PCR analysis of *Reptin* (primers F1, R) **(B)** Relative Reptin mRNA expression level from liver (n=5), heart, testis, kidney, lung, muscles, brain, spleen (n=3) and pancreas (n=2) extracts from *Reptin*<sup>fl/fl</sup> and *Reptin*<sup>LKO</sup> mice D15 post-TAM. **(C)** Plasma ALT and AST measurements (n=3). **(D)** Representative pictures of BrdU and active-caspase 3 immunostaining of liver sections from *Reptin*<sup>fl/fl</sup> and *Reptin*<sup>LKO</sup> mice at D15 post-TAM. Scale bar, 100  $\mu$ m. **(E)** Body mass gain calculation and **(F)** Relative mRNA expression of Reptin in liver extracts from *Reptin*<sup>fl/fl</sup> and *Reptin*<sup>LKO</sup> mice from D0 to 6 weeks post-tamoxifen injection (n=3 per group). **(G)** Total meal per day in mice fed ad libitum; **(H)** Size and number of meals per day and; **(I)** Food intake ad libitum and during fasting and refeeding as indicated in *Reptin*<sup>fl/fl</sup> and *Reptin*<sup>LKO</sup> mice (n=6). **(J)** Fuel oxidation ratio from *Reptin*<sup>fl/fl</sup> and *Reptin*<sup>LKO</sup> mice during refeeding (n=6). **(K)**  $\beta$ -hydroxybutyrate plasma levels from *Reptin*<sup>fl/fl</sup> and *Reptin*<sup>LKO</sup> mice at D15 post-TAM fed AL (n=5). All graphs represent mean  $\pm$  SEM and significance is indicated as follows: \*  $p \leq 0.05$ , \*\*  $p \leq 0.01$ .



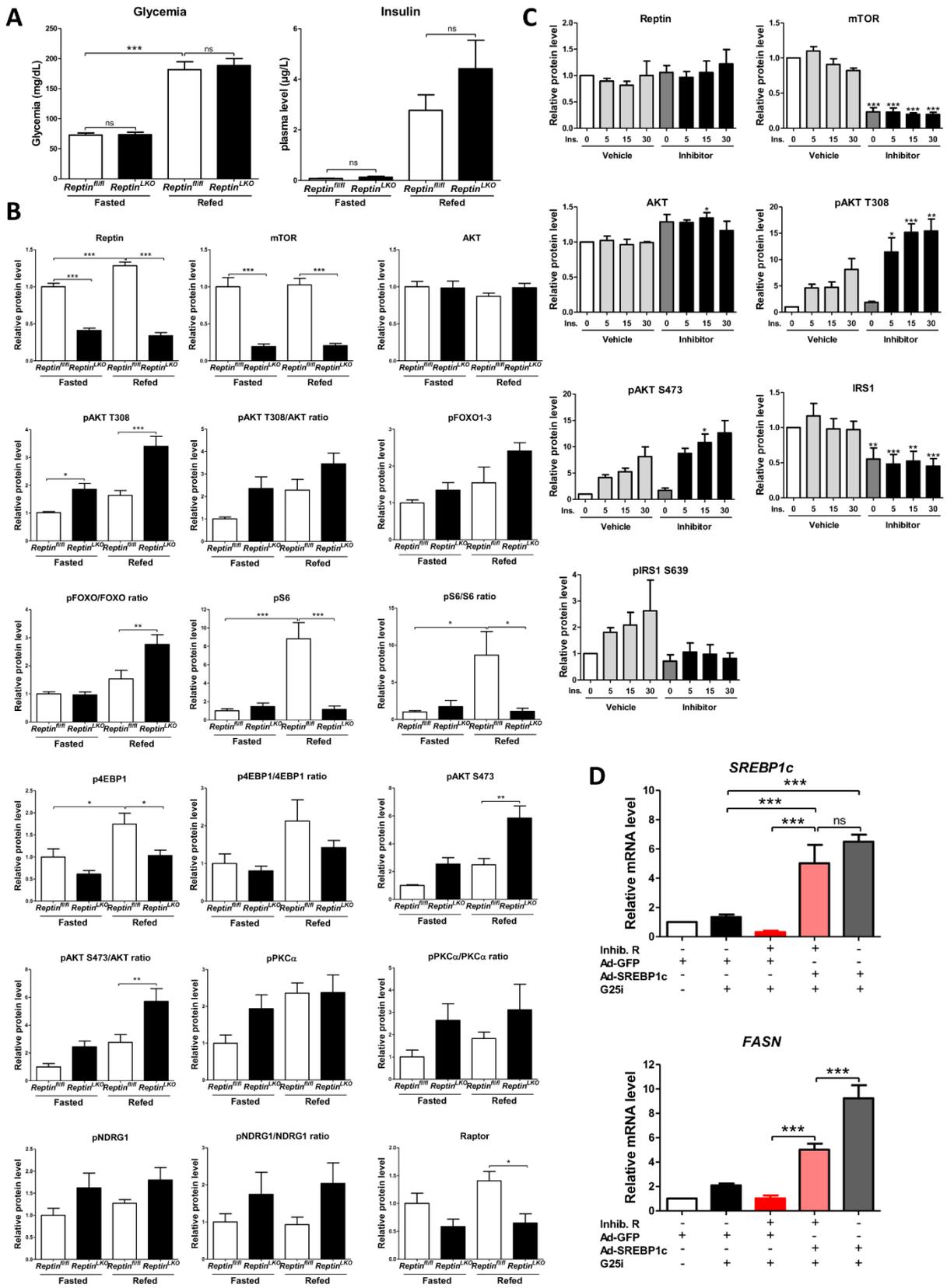
**Sup Figure 2: Loss of hepatic Reptin leads to decreased mTOR protein level in a cell-autonomous-manner**

**(A)** Representative immunoblotting and relative protein expression of Reptin and mTOR and **(B)** relative mRNA expression of mTOR in *Reptin*<sup>lox/lox</sup> mouse primary hepatocytes extracts after infection with Cre- or GFP-expressing adenovirus at MOI 10 (n=2). **(C)** mTOR immunofluorescence in *Reptin*<sup>lox/lox</sup> mouse primary hepatocytes infected with Cre- or GFP-expressing adenovirus at MOI 10. Scale bar, 20  $\mu$ m. All graphs represent mean  $\pm$  SEM. **(D)** Representative immunoblotting of mTOR, DNA-PKcs and ATR in primary hepatocytes treated with Reptin/Pontin ATPase inhibitor (1 $\mu$ M) or vehicle (DMSO) (n=3). DNA-PKcs and ATR are other members of the PIKK family also known to be chaperoned by the R2TP complex (6) **(E)** Sulforhodamine-B cytotoxicity assay in primary hepatocytes treated with Reptin/Pontin ATPase inhibitor at indicated concentrations or vehicle (DMSO). Etoposide has been used as a positive control. **(F)** Representative immunoblotting of mTOR in primary hepatocytes treated with Reptin/Pontin ATPase inhibitor (1 $\mu$ M). Cells were then treated with proteasome inhibitors MG132 (10  $\mu$ M) or Bortezomib (10  $\mu$ M) 9 hours before harvest. The accumulation of phosphorylated  $\beta$ -catenin demonstrates the efficiency of proteasome inhibition. Tubulin was used as loading controls.

**A****B****C****D****E****F**

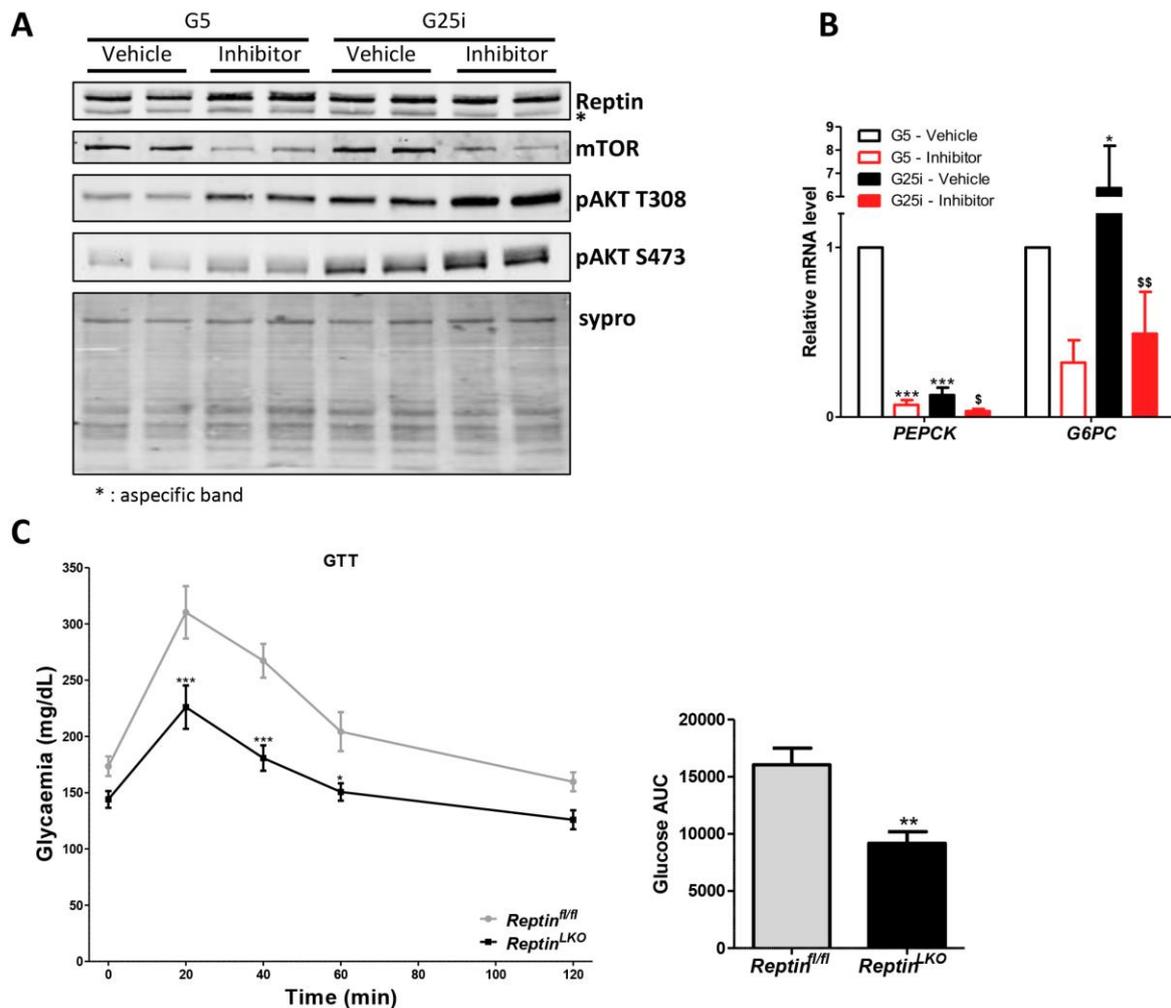
**Sup Figure 3: Opposite effects on mTORC1 and mTORC2 signaling upon loss of hepatic Reptin.**

**(A)** Glycemia and plasma insulin levels measured upon 13 hours fasting and 6 hours refeeding in *Reptin<sup>fl/fl</sup>* and *Reptin<sup>LKO</sup>* mice. **(B)** Relative protein expression of Reptin, mTOR, AKT, mTORC1 and mTORC2 signaling in liver extracts from *Reptin<sup>fl/fl</sup>* and *Reptin<sup>LKO</sup>* mice in fasted and refed state (n=6, from 2 independent experiments). **(C)** Relative protein expression of insulin signaling in mouse primary hepatocytes extracts after treatment with 1  $\mu$ M Reptin/Pontin ATPase inhibitor and stimulation by 10 nM insulin as shown on the graph (n=3). In these experiments, significance is shown between the conditions vehicle and Reptin ATPase inhibitor at every time point. **(D)** Relative mRNA levels of *DNL genes SREBP1c and FASN* in primary hepatocytes treated with Reptin/Pontin ATPase inhibitor (1 $\mu$ M) and transduced with Ad-SREBP1c active or Ad-GFP as control (MOI 1). Cells were stimulated with glucose 25mM and insulin 100 nM for 24h (n=4). All graphs represent mean  $\pm$  SEM and significance is indicated as follows: \*  $p \leq 0.05$ , \*\*  $p \leq 0.01$ , \*\*\*  $p \leq 0.001$ , ns (not significant).



**Sup Figure 4: Loss of Reptin in the liver improves glucose tolerance.**

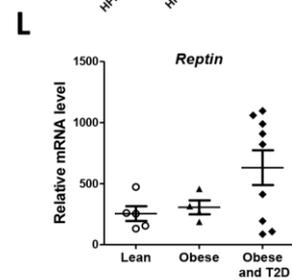
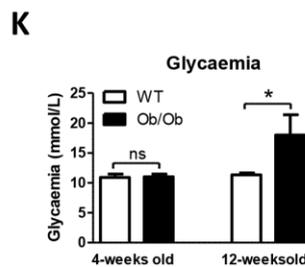
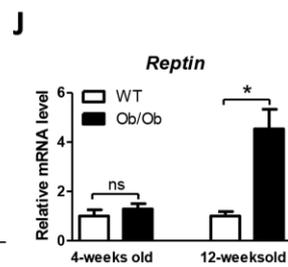
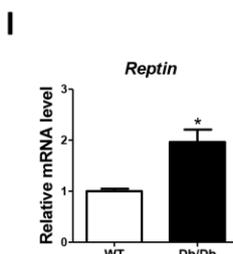
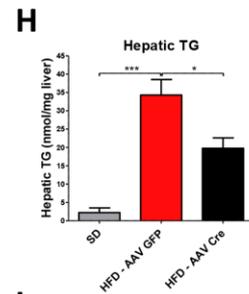
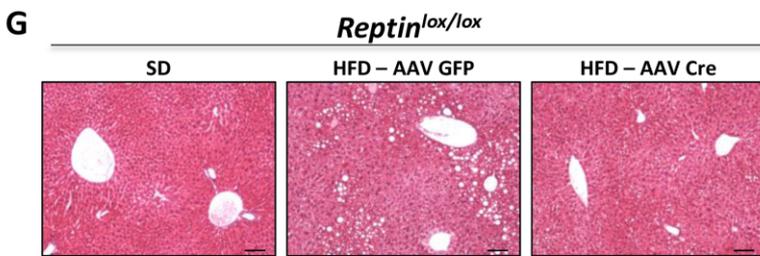
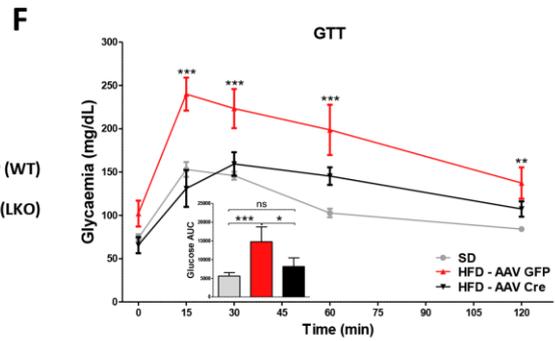
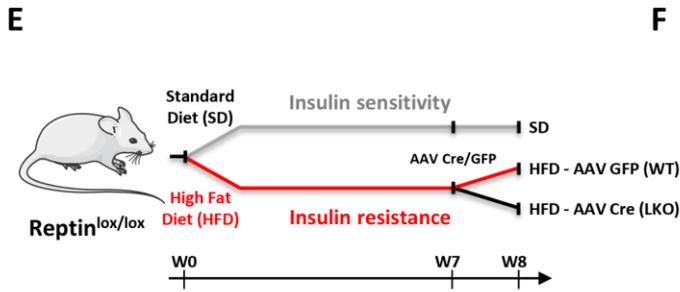
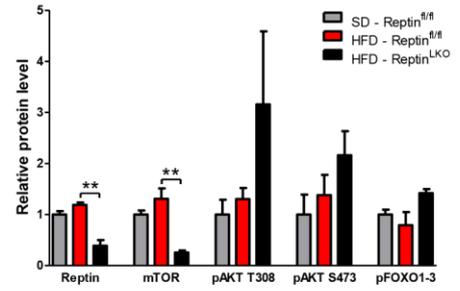
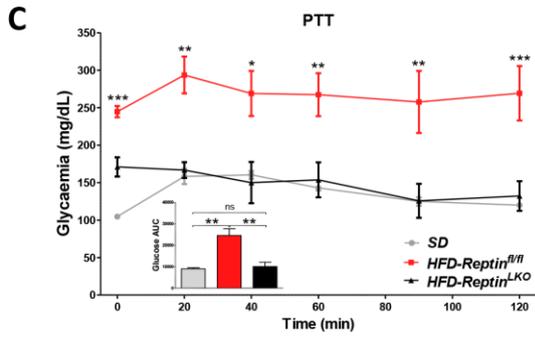
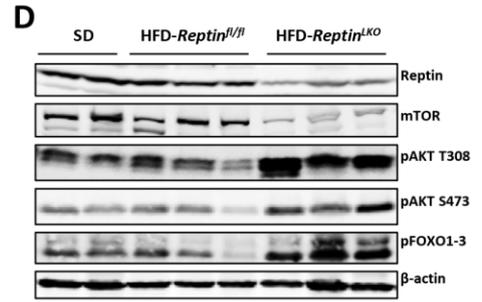
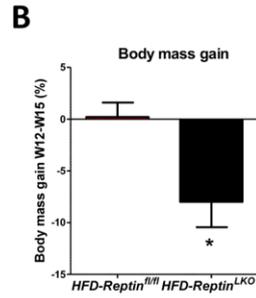
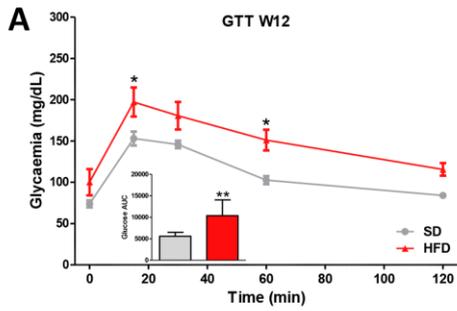
**(A)** Representative immunoblotting of mTOR and pAKT Ser473 and Thr308 and **(B)** relative mRNA expression of gluconeogenic genes (*PEPCK* and *G6PC*) genes expression in mouse primary hepatocytes extracts stimulated for 24h with 5 mM (G5) or with 25 mM glucose and 100 nM insulin (G25i) after treatment with 1  $\mu$ M Reptin ATPase inhibitor or control-Vehicle (n=3). All graphs represent mean  $\pm$  SEM; significance versus 5 mM low glucose control (G5-Vehicle) is indicated as follows: \* p $\leq$ 0.05, \*\* p $\leq$ 0.01, \*\*\* p $\leq$ 0.001 and significance versus 25 mM high glucose + 100 nM insulin control (G25i-Vehicle) is indicated as follows: \$ p $\leq$ 0.05, \$\$ p $\leq$ 0.01, \$\$\$ p $\leq$ 0.001. **(C)** Oral Glucose Tolerance Test (1g/kg) with quantification of the area under the curve from *Reptin*<sup>fl/fl</sup> and *Reptin*<sup>LKO</sup> mice after 8 hours of fasting (n=7). All graphs represent mean  $\pm$  SEM and significance is indicated as follows: \* p $\leq$ 0.05, \*\* p $\leq$ 0.01, \*\*\* p $\leq$ 0.001.



### Sup Figure 5: Loss of Reptin rescues HFD-induced insulin resistance and hepatic steatosis

**(A)** Oral Glucose Tolerance Test (1g/kg) after 12 hours of fasting with quantification of the area under the curve from SD (n=3) and HFD (n=8) mice 12 weeks after HFD introduction and prior induction of Reptin deletion. Significance is shown between SD and HFD at every time point. **(B)** Body weight gain calculation from HFD-*Reptin<sup>fl/fl</sup>* (n=3) and HFD-*Reptin<sup>LKO</sup>* (n=5) mice 18 days after tamoxifen injection. **(C)** Hepatic glucose production analysis by Pyruvate Tolerance Test (PTT) 2g/kg after 10 hours of fasting with quantification of the area under the curve calculated from baseline  $y=100$  from SD (n=3), HFD-*Reptin<sup>fl/fl</sup>* (n=3) and HFD-*Reptin<sup>LKO</sup>* (n=5) mice 16 days after TAM injection. Significance is shown between SD and HFD-*Reptin<sup>fl/fl</sup>* mice at every time point. **(D)** Representative immunoblotting of insulin signaling in liver extracts from SD, HFD-*Reptin<sup>fl/fl</sup>* and HFD-*Reptin<sup>LKO</sup>* 18 days after Reptin deletion. **(E)** Experimental strategy for Reptin deletion by retro-orbital injection of Cre-expressing AAV in *Reptin<sup>lox/lox</sup>* model with 8 weeks HFD-induced metabolic syndrome. Three groups: standard diet (SD n=3), HFD-AAV GFP (n=3) and HFD-AAV Cre (n=3). **(F)** Glucose tolerance was restored back to normal in mice with AAV-Cre Reptin-deficient livers. **(G)** Hepatic steatosis was abolished as revealed by H&E staining of liver sections (scale bar, 100  $\mu$ m) and **(H)** hepatic TG content was decreased in AAV-Cre Reptin-deficient livers. Significance is shown between SD and HFD-AAV GFP mice at every time point. **(I-J)** Relative mRNA of Reptin in liver extracts of **(I)** diabetic Db/Db mice (n=5), and **(J)** of 4- and 12- weeks- old obese Ob/Ob mice (n=3) with **(K)** respective glycemia levels (n=3). All graphs represent mean  $\pm$  SEM and significance is indicated as follows: \*  $p \leq 0.05$ , \*\*  $p \leq 0.01$ , \*\*\*  $p \leq 0.001$ . **(L)** Reptin mRNA level in liver of obese or obese and T2D subjects compared to lean subjects (7).

**Data from ME Patti's lab, Boston (GEO analysis):** <https://www.ncbi.nlm.nih.gov/gate2.inist.fr/geo/query/acc.cgi?acc=GSM391710>; [https://www.ncbi.nlm.nih.gov/gate2.inist.fr/geo/tools/profileGraph.cgi?ID=GDS3876:201459\\_at](https://www.ncbi.nlm.nih.gov/gate2.inist.fr/geo/tools/profileGraph.cgi?ID=GDS3876:201459_at).



**Sup Figure 6: Loss of Reptin increases mTORC2 signalling.**

**(A)** Relative mRNA expression of *INSIG2A* in liver extracts from *Reptin<sup>fl/fl</sup>* and *Reptin<sup>LKO</sup>* mice as indicated (n=6, from 2 independent experiments). **(B)** Venn diagrams of UP and DOWN-regulated genes that are affected both in fasted and refed conditions in liver extracts from *Reptin<sup>LKO</sup>* mice. **(C)** Relative mRNA expression of *Rictor* in liver extracts from *Reptin<sup>fl/fl</sup>* and *Reptin<sup>LKO</sup>* mice as indicated (n=6, from 2 independent experiments). All graphs represent mean ± SEM and significance is indicated as follows: \* p≤0.05, \*\* p≤0.01, \*\*\* p≤0.001, ns (not significant).

