OPEN ACCESS

ORIGINAL ARTICLE

Scientific frontiers in faecal microbiota transplantation: joint document of Asia-Pacific Association of Gastroenterology (APAGE) and Asia-Pacific Society for Digestive Endoscopy (APSDE)

Siew C Ng ,^{1,2} Michael A Kamm,³ Yun Kit Yeoh ,^{1,4} Paul K S Chan,^{1,4} Tao Zuo,^{1,2} Whitney Tang,^{1,2} Ajit Sood,⁵ Akira Andoh,⁶ Naoki Ohmiya,⁷ Yongjian Zhou,⁸ Choon Jin Ooi,⁹ Varocha Mahachai,^{10,11} Chun-Ying Wu,^{12,13} Faming Zhang,^{14,15} Kentaro Sugano,¹⁶ Francis K L Chan ^{1,2}

► Additional material is published online only. To view please visit the journal online (<http://dx.doi.org/10.1136/gutjnl-2019-319407>).

For numbered affiliations see end of article.

Correspondence to

Professor Siew C Ng, Center for Gut Microbiota Research, Chinese University of Hong Kong, Hong Kong, China; siewchienng@cuhk.edu.hk

Received 17 July 2019

Revised 6 September 2019

Accepted 24 September 2019

Published Online First

14 October 2019

ABSTRACT

Objective The underlying microbial basis, predictors of therapeutic outcome and active constituent(s) of faecal microbiota transplantation (FMT) mediating benefit remain unknown. An international panel of experts presented key elements that will shape forthcoming FMT research and practice.

Design Systematic search was performed, FMT literature was critically appraised and a 1-day round-table discussion was conducted to derive expert consensus on key issues in FMT research.

Results 16 experts convened and discussed five questions regarding (1) the role of donor and recipient microbial (bacteria, viruses, fungi) parameters in FMT; (2) methods to assess microbiota alterations; (3) concept of keystone species and microbial predictors of FMT, (4) influence of recipient profile and antibiotics pretreatment on FMT engraftment and maintenance and (5) new developments in FMT formulations and delivery. The panel considered that variable outcomes of FMT relate to compositional and functional differences in recipient's microbiota, and likely donor-associated and recipient-associated physiological and genetic factors. Taxonomic composition of donor intestinal microbiota may influence the efficacy of FMT in recurrent *Clostridioides difficile* infections and UC. FMT not only alters bacteria composition but also establishes trans-kingdom equilibrium between gut fungi, viruses and bacteria to promote the recovery of microbial homeostasis. FMT is not a one size fits all and studies are required to identify microbial components that have specific effects in patients with different diseases.

Conclusion FMT requires optimisation before their therapeutic promise can be evaluated for different diseases. This summary will guide future directions and priorities in advancement of the science and practice of FMT.

BACKGROUND

Faecal microbiota transplantation (FMT) has been shown to be a highly efficacious, safe and cost-effective therapy for recurrent *Clostridioides difficile* infections (CDI).^{1,2} Beyond CDI, FMT has

Significance of this study**What is already known on this subject?**

- Faecal microbiota transplantation (FMT) is effective in recurrent *Clostridioides difficile* infections and is potentially beneficial in other microbiota-related disorders.
- FMT rectifies the recipient's intestinal microbial community by introducing micro-organisms associated with a 'healthy' state to normalise microbiota composition and function.

What are the new findings?

- Variable outcomes of FMT may relate to compositional and functional differences in the recipient's gut microbiota, and likely donor-associated and recipient-associated physiological, genetic or demographic factors.
- Taxonomic composition of donor intestinal microbiota may influence the efficacy of FMT in recurrent *Clostridioides difficile* infections and UC.
- FMT helps establish trans-kingdom equilibrium between gut fungi, viruses and bacteria to promote the recovery of microbial homeostasis.
- FMT is not a one size fits all strategy, and studies are required to identify microbial components microbiota that have specific effects in patients with different diseases.

How might it impact on clinical practice in the foreseeable future?

- Selecting donors based on microbial indicators and/or capability of the donor microbiota to restore specific metabolic disturbances associated with each disease may be important for improved FMT efficacy.
- FMT delivery methods and microbial strains need to be rationally designed and tightly controlled taking into account microbial diversity, taxonomic composition and donor-recipient factors.

© Author(s) (or their employer(s)) 2020. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.

To cite: Ng SC, Kamm MA, Yeoh YK, et al. *Gut* 2020;**69**:83–91.

been investigated as a treatment option in a variety of diseases (GI), including IBD,^{3–6} IBS,⁷ hepatic encephalopathy,^{8,9} autism¹⁰ and metabolic syndrome.^{11,12} Although FMT is potentially effective, substantial heterogeneity in donor selection and FMT application have been reported in studies across different populations.¹³ According to data submitted to the clinical studies registry in the US National Institutes of Health, over 200 clinical trials of FMT in various diseases are being conducted or have been registered in the past 2 years.¹⁴ However, the underlying microbial predictors of therapeutic outcome and active constituent(s) of FMT mediating success remain largely unknown, even in the setting of the most studied recurrent CDI (rCDI). Recently, the term ‘super-donor’ has been proposed to describe donors whose stool results in more successful FMT outcomes compared with stool from ‘regular’ donors.¹⁵ This paper aims to review the scientific rationale behind FMT with a main focus on recent advances in our understanding of mechanisms of FMT and factors associated with FMT outcomes across different clinical conditions. It does not intend to cover clinical aspects of FMT, such as indications or outcome, donor screening^{2,16} or regulations of FMT practice,^{17,18} as most of these aspects have been extensively reviewed elsewhere. In a frontiers meeting held in Hong Kong, experts within Asia-Pacific presented their views and discussed outlooks pertaining to FMT regarding (1) the role of donor and recipient microbial parameters; (2) concept of keystone species and microbial predictors, (3) methods to assess microbial alterations, (4) possible influence of recipient profile and antibiotics pretreatment on FMT engraftment and maintenance and (5) new developments in FMT formulations and delivery. Detailed methodology is described in online supplementary appendix 1 and figure 1 shows the flow diagram of studies included.

MICROBIOLOGY OF FMT

Role of donor-associated microbial parameters in FMT

Recurrent CDI

While current donor screening procedures mainly focus on safety, by excluding known human pathogens, there is no clear agreement on selecting donors based on microbial parameters.¹⁶

In a study where 10 paediatric patients with rCDI were treated with FMT from a competitively selected ‘optimal’ universal donor based on high microbial diversity, balanced Bacteroidetes versus Firmicutes and a high concentration of faecal butyrate, all subjects showed no disease recurrence at 10 weeks after treatment.¹⁹ This study demonstrated that using markers of the bacterial community profile may be feasible for donor stratification, although larger cohorts inclusive of control populations without specific donor profiling are needed to confirm whether donor stratification improves FMT success rates in rCDI.

Few studies have focused on potential role of viruses as a mediator of FMT efficacy. This is due to complexities in profiling viruses compared with bacteria as viruses lack universal marker genes. To facilitate recovery of virus sequences in metagenomes, virome-enrichment nucleic acid extraction methods have been developed to selectively enrich virus particles from stool. However, these enrichment procedures disrupt inherent virus community by altering their relative numbers. One pilot study of five subjects administered a single jejunal infusion of sterile stool filtrate and all subjects with relapsing CDI recovered, lending support to a role of bacteriophages in FMT efficacy.²⁰ The positive outcome from using non-live bacterial stool filtrates in FMT highlighted the possibility that bacterial fractions of donor gut communities are necessary for successful treatment, although

Figure 1 Flow diagram of studies included after systematic search.

it did not exclude that bacterial-secreted metabolites and cell components such as proteins and RNAs that may exert crucial effects in the recipient. In a survey of enteric bacterial and viral composition in an FMT recipient cured of rCDI, the recipient’s phage community was highly similar to the donor’s following FMT and remained stable for 7 months.²¹ The viral communities in both donor and post-transplant recipient exhibited comparable diversity and richness. These studies indicate that enteric virome composition could be a key indicator of the clinical outcomes of FMT for rCDI.²¹ We found that in rCDI patients treated with FMT, all recipients responded to FMT when richness of Caudovirales was higher in donors compared with recipients.²² A separate study of 19 patients with rCDI showed that subjects successfully treated with a single FMT had increased Bacteroidetes and Firmicutes and a complete loss or reduction of Siphoviridae phages. FMT can alter both bacterial and phage composition to resemble the donor.²³ Patients requiring multiple FMT showed increased abundances of *Enterobacteria* phages HK542, mEp237 and phiP27 suggesting that the presence of these *Enterobacteria* phages may be associated with reduced efficacy.²³

The human gut also hosts various fungi and other eukaryotes.²⁴ Fungal communities are commonly profiled using a DNA barcode sequence known as the internal transcribed spacer (ITS),²⁵ but this approach has multiple shortcomings including large variation in copy number among fungal taxa, variable lengths in ITS sequences and fungal cells are harder to lyse during DNA extraction. These factors, combined with a significantly less comprehensive reference database compared with the 16S ribosomal RNA gene, distort relative abundance estimates in fungal community surveys. It was found that higher relative abundances of *Saccharomyces* and *Aspergillus* in stool of recipients correlated with successful FMT for rCDI and a

high abundance of *Candida albicans* in donor stool was associated with reduced FMT efficacy.²⁶ Both gut bacteriophages and fungi, particularly *C. albicans*, affect bacterial community assembly,^{27–29} potentially influencing FMT effectiveness in rCDI.

Inflammatory bowel disease

In IBD, a small study of 14 refractory patients (8 UC, 6 Crohn's disease (CD)) reported that higher bacterial species richness in donors and a higher number of transferred phylotypes were associated with successful transplantation.³⁰ In addition, stool of donors with a high bacterial richness, a high relative abundance of *Akkermansia muciniphila*, unclassified Ruminococcaceae and *Ruminococcus* spp were more likely to induce remission.³¹ In a randomised controlled trial (RCT) of FMT in UC, increased relative abundances of *Eubacterium hallii* and *Roseburia inulinivorans* in UC were the strongest and most consistent associations with positive clinical outcomes. Functional analyses demonstrated that *Eubacterium* and *Roseburia* contributed to increases in beneficial metabolic pathways.³² For instance, *E. hallii* maintains microbial metabolic health and homeostasis via short-chain fatty acid production. In contrast, the presence of *Fusobacterium* in UC was associated with FMT failure.³² Reduced relative abundances of *Clostridium* cluster XIVa and increased Bacteroidetes were reversed by FMT, mainly in responders.³³ Sustained remission was associated with butyrate-producing organisms, and relapse was associated with Proteobacteria and Bacteroidetes.³³ It has also been suggested that remission rates could be enhanced by pooling stools from multiple donors to increase microbial diversity.^{5,34}

A low eukaryotic virus richness before FMT in patients with UC was associated with FMT success.³⁵ These studies showed that phages in donor stool were associated with FMT efficacy in CDI, whereas eukaryotic viruses appeared to be more relevant in UC, suggesting that virome interactions may be disease specific. Although phages may play a role in efficacy, the experimental designs of many studies did not exclude the possibility that these viruses were purely enriched alongside bacterial hosts, as whole donor stools including bacteria were transplanted into recipients.

Other diseases

With improvements in high-throughput DNA sequencing technology, it is now possible to study long-term engraftment of donor bacterial strains.³⁶ In a study into metabolic syndrome, multiple recipients of a single donor showed extensive coexistence of donor and recipient faecal bacterial strains up to 3 months after FMT. Furthermore, recipients receiving the same donor stool displayed large variation in microbiota transfer, indicating that individuals vary in their microbiome engraftment and donor–recipient compatibility.³⁷ These examples indicate that strain level differences between microbial taxa could offer a key to understanding why certain strains are more efficacious relative to others, and that these genomic differences can be used to optimise donor selection. Recent work has evaluated the temporal variation in bacteriome and virome after FMT. In a small pilot study of three patients with carbapenem-resistant Enterobacteriaceae who had successful decolonisation after FMT, viral communities varied substantially over time whereas intensive trans-kingdom correlations between viruses (Caudovirales) and bacteria (Ruminococcaceae) were significantly depleted in recipients after FMT, particularly with the interaction between *Klebsiella*-specific bacteriophages and Ruminococcaceae species weakened.³⁸ In acute graft versus host disease

(GvHD), a series of FMTs altered the gut bacterial, fungal and viral communities in a patient with GvHD. Engraftment of donor-derived fungi occurred instantly after a single FMT and persisted up to 4 months³⁹. Moreover, FMT reduced an over-represented fungus *Fusarium oxysporum* and Torque teno virus in parallel with an increase in fungal diversity and abundance of Caudovirales bacteriophages. While these data do not directly address the role of fungi in FMT, it suggests that FMT helps establish trans-kingdom equilibrium between gut fungi, viruses and bacteria to promote recovery.³⁹ Although the clinical value of donor selection based on stool microbial community profiles remains to be established, incorporation of steps to assess stool community composition is technically feasible. Since microbial community composition, richness and diversity remained a distinct characteristic of each donor, a single assessment of stool composition during donor selection is likely sufficient to represent samples collected in subsequent months.⁴⁰

Table 1 summarises donor and recipient stool compositions and factors associated with improved FMT efficacy.

Microbial predictors of FMT success: how to study them

To understand how gut microbial communities are involved in FMT and whether key micro-organisms can be manipulated to improve therapeutic outcomes, studies have used 16S ribosomal RNA gene-based amplicon sequencing to characterise the gut microbiota of FMT donors and recipients. In successful cases of treating UC and rCDI, general trends observed in the recipients' gut community profiles are the following: (1) increased bacterial community diversity following FMT,^{3,41,42} (2) community profiles more closely resembling donor profiles,^{41–43} (3) decrease in certain bacterial taxa including Proteobacteria and Fusobacteria and (4) a corresponding increase in key Firmicutes and Bacteroidetes taxa.^{33,41–44} In patients with metabolic syndrome, lower initial faecal microbiota diversity before FMT in the recipients was predictive of metabolic improvement on treatment.^{11,12} In addition to donor-derived microbial populations that persist in the recipient, some microbial populations that flourish post-FMT are undetectable in donor or recipient pretransplantation. Taken together, these findings indicate that micro-organisms from donors engraft in recipients, potentially colonising and/or creating conditions that repress disease-associated micro-organisms. This change in the gut environment could create conditions that allow microbial populations below detection thresholds in pre-FMT samples to flourish and repopulate the gut, helping to restore diseased gut communities to a 'healthy' state. Although most FMT studies have used 16S rRNA profiling, 16S sequences do not contain sufficient resolution to inform whether the same microbial species or strains are found in both donors and recipients. An increasing number of studies now use shotgun metagenomics (sequencing of bulk DNA) in which total extracted DNA is sequenced without preselection for any marker genes. Given the strong influence of environment on human gut microbiota composition⁴⁵ and a high functional redundancy among gut microbial taxa,⁴⁶ it is highly likely that FMT success is mediated by functionality of gut micro-organisms in addition to phylogeny. Using *gene-centric metagenomics*, community functionality is traditionally assessed by sequencing bulk DNA and then comparing the resultant sequences to gene orthology databases such as the Clusters of Orthologous Groups⁴⁷ and Kyoto Encyclopedia of Genes and Genomes.⁴⁸ Genome-centric metagenomics has been widely used in environmental microbiology but is not yet commonplace in human gut microbiome studies.^{49,50} To date, the use of

Table 1 Donor and recipient stool compositions and factors associated with improved FMT efficacy

Condition	Criteria of donor stool	Criteria of recipient stool
<i>Clostridioides difficile</i> infection	<ul style="list-style-type: none"> ▶ High microbial bacteria diversity¹ ▶ Balanced constitution of Bacteroidetes versus Firmicutes¹ ▶ High concentration of faecal butyrate¹ ▶ Caudovirales richness in donor stool higher than recipient stool² ▶ Low abundance of <i>C. albicans</i>³ 	<p>Before FMT</p> <ul style="list-style-type: none"> ▶ Higher relative abundances of <i>Saccharomyces</i> and <i>Aspergillus</i>³ ▶ Low abundance of <i>Candida albicans</i>³ ▶ Low abundance of <i>Enterobacteria</i> phages⁴ <p>After FMT</p> <ul style="list-style-type: none"> ▶ Increase in donor-derived phages⁴ ▶ Complete loss or significant reduction of Siphoviridae⁴ ▶ Increased relative abundance of Bacteroidetes and Firmicutes⁴ ▶ Low Enterobacteriaceae⁵
UC	<ul style="list-style-type: none"> ▶ High bacteria species richness and diversity^{6,7} ▶ Low abundance of <i>Fusobacterium</i>⁸ ▶ High relative abundance of <i>Akkermansia muciniphila</i>, unclassified Ruminococcaceae and <i>Ruminococcus</i> spp⁷ ▶ Low level of <i>Ruminococcus gnavus</i>⁸ 	<p>Before FMT</p> <ul style="list-style-type: none"> ▶ Low eukaryotic viral richness⁹ ▶ Absence of <i>Fusobacterium</i>¹⁰ <p>After FMT</p> <ul style="list-style-type: none"> ▶ Increased microbial diversity⁷ ▶ Increased <i>Clostridium</i> cluster IV (eg, <i>Faecalibacterium prausnitzii</i>) and XIVa (<i>Eubacterium hallii</i>, <i>Roseburia intestinalis</i>, <i>Roseburia inulinivorans</i>, <i>Butyrivibrio crossotus</i>)^{8,11} ▶ Reduction of Proteobacteria (<i>Escherichia coli</i> or <i>Aeromonas</i>) and Bacteroidetes⁸
Metabolic syndrome	NA	<p>Before FMT</p> <ul style="list-style-type: none"> ▶ Lower initial faecal microbiota diversity¹² ▶ Higher relative abundances of <i>Subdoligranulum variabile</i> and <i>Dorea</i>¹² ▶ Lower <i>Eubacterium ventriosum</i> and <i>Ruminococcus torques</i>¹²
IBS	▶ High abundance of <i>Bifidobacterium</i> ¹³	NA

Refer to Appendix 2 for references (online supplementary file 1)

FMT, faecal microbiota transplantation; NA, not applicable.

shotgun metagenomics screening to predict FMT success in CDI and other diseases is mostly confined to academic research due to its cost and timescale. Nevertheless, information gained from this research can be instrumental in formulating clinical trials, such as guiding primer designs for screening a list of beneficial and negative micro-organisms and viruses. Table 2 illustrates the different methods to characterise microbial parameters in FMT. Figure 2 illustrates the potential mechanism of FMT.

Clinical predictors of FMT

Several clinical factors have been reported to be associated with failure of FMT in rCDI. In a prospective study of 64 patients with rCDI treated with FMT by colonoscopy, severe CDI and inadequate bowel preparation were independent predictors of failure after single faecal infusion.⁵¹ A multicentre study showed that predictors of early FMT failure included severe or severe-complicated CDI, inpatient status during FMT and previous CDI-related hospitalisation were associated with early FMT failure.⁵² In a RCT of patients with active UC, those with disease for less than 1 year appeared to have better outcomes after FMT.³

OPTIMISING FMT PROTOCOLS

Donor–recipient matching

It is recognised that differences in microbiome composition between individuals and diseases could contribute to variations in response to FMT.⁵³ Furthermore, host genetic factors such as differences in innate immune responses or environmental factors, such as diet, xenobiotic exposure and microbial interactions may also play a role in shaping FMT responses between donor and recipient.¹⁵ OpenBiome, a public stool bank in the USA published an investigation into the selection criteria of universal donors based on faecal butyrate concentration, microbial diversity metrics and ratio of Firmicutes to Bacteroidetes which yielded good FMT outcomes for rCDI, but there was no formal comparison of these selection criteria against standard

donor selection whereby microbiota composition or metrics were not taken into account for FMT.¹⁹ Donor–recipient matching for FMT may not be as critical for CDI because of high efficacy even without specific matching, but is likely important for other conditions including UC and IBS in which response rates remain modest and variable (approximately 30%–40%).^{7,54} For example, when gut microbiota profiles of different donors were compared, it was found that microbial diversity was a predictor for FMT success in UC.³¹ While current evidence suggests that FMT efficacy can be improved by carefully selecting donors based on their gut microbiota content and that ‘one stool does not fit all’, the specific parameters for implementing a selection criteria are still unknown. Alterations in microbiota may also not be the only factor contributing to disease status or development. An individual’s response to FMT may predominantly depend on the capability of the donor’s microbiota to engraft and reverse the microbial community dysbiosis associated with a specific disease phenotype; these factors need to be investigated in future studies.

Apart from using whole faecal transplants, one approach which removes donor-dependent effects is to provide patients with a defined cocktail of micro-organisms confirmed to be beneficial for disease resolution. For example, providing IBD patients with a targeted microbiota-based formulation containing only butyrate producers may be a logical and safer alternative to whole faecal transplantation. Precision approaches have been trialled in CDI but with mixed results.^{55,56} It may be that the donor microbial community as a whole plays a stronger role in driving FMT success than critical species in isolation. Nevertheless, targeted bacteriotherapy approaches should be investigated as a way of circumventing risks associated with administering whole faecal material.

Recipient microbial profiling prior to FMT

The criteria that denote an ‘optimal recipient’ for receiving FMT are not clear, although there are several studies describing

Table 2 Methods to study microbial parameters in FMT

Types of sequencing	Details of methods
16S rRNA profiling	<ul style="list-style-type: none"> ▶ 16S rRNA gene-based amplicon sequencing is less costly and is commonly used to characterise the gut microbiota of FMT donors and recipients ▶ 16S sequences do not contain sufficient resolution to inform whether the same microbial species or strains are found in both donors and recipients
Shotgun metagenomics	<ul style="list-style-type: none"> ▶ Breadth of sequences covering entire genomes provides additional information, such as gene content and nucleotide polymorphisms, to more accurately infer movement of microbial populations from donor to recipient. ▶ Assembly of DNA sequences into contiguous sequences and reconstruction of bacteria genomes (termed <i>binning</i>) can be used to identify exact microbial strains transferred between donor and recipients and their genetic potential. ▶ Ability to compare genome content across studies, assessment of metabolic functions to predict engraftment, and tracking the transfer of microbial genomes in pooled multi-donor FMTs
Gene-centric metagenomics	<ul style="list-style-type: none"> ▶ Sequence counts are used as a proxy for functionality.¹⁴ ▶ There are few metagenome-based studies of FMT gut communities,^{15–17} but none specifically compares gene content between donors and recipients before and after FMT. For example, increased concentrations of gamma-aminobutyric acid in plasma,¹² short-chain fatty acids¹⁸ and secondary bile acids¹⁹ have been demonstrated in recipients following FMT for CDI. ▶ Gene sequence data can be verified against measurable parameters to confirm whether metagenomics data corroborates changes in metabolites associated with FMT, and provide support for (or against) the use of DNA sequencing in studying mechanisms of FMT.
Genome-centric metagenomics	<ul style="list-style-type: none"> ▶ Genome-centric metagenomics aims to reconstruct microbial population genomes from sequence data (<i>binning</i>) without relying on reference genomes.¹⁵ ▶ Genomes recovered this way, commonly termed metagenome-assembled genomes or MAGs, represent sample-specific microbial populations and can be analysed for their gene sequence and content, metabolic potential, correlated with patient-specific clinical data or serve as mapping references for downstream transcriptomic studies. <p>Examples of studies using this approach:</p> <ul style="list-style-type: none"> ▶ This approach has been applied in two separate studies to identify the donor microbial populations that establish in recipients. The authors reported establishment by Bacteroidetes and Firmicutes taxa.^{15,17} Lee and colleagues reconstructed population genomes from donor and recipient metagenomes and used read mappings to detect their presence across subjects.¹⁵ Using a slightly different approach, Smillie and colleagues mapped metagenome sequence reads to reference microbial genomes from the Human Microbiome Project and inferred single-nucleotide polymorphisms in read alignments to detect strains shared by donors and recipients post-FMT.¹⁷ Both studies reported variability in the outcomes of donor micro-organisms engrafting in recipients, with different species or strains showing establishment in some recipients and not others. The amount of engraftment also varied, with donor strains constituting between 1% and 80% of the recipient gut community.¹⁷ Smillie and colleagues built a statistical model based on their data to predict patterns of engraftment, reporting that bacterial taxonomy, abundance in donors/recipients and elapsed time post-FMT were the major determining factors.¹⁷ Some donor-derived strains could be detected in recipients for only 1 day post-FMT, and others persisted for longer than 1 month.¹⁷ Although the exact ecological drivers for engraftment and persistence are not yet resolved, it was demonstrated that it is possible to derive genomic information with enough resolution to track microbial engraftment in FMT through metagenomics.

Refer to Appendix 2 for references (online supplementary file 1)
FMT, faecal microbiota transplantation; rRNA, ribosomal RNA.

correlations between gut microbiota composition of recipients and FMT outcomes. In a study of patients with UC receiving FMT, the presence of *Fusobacterium* in recipient stool was associated with FMT failure.³² In another study of subjects with metabolic syndrome, responders had lower initial faecal microbiota diversity, higher relative abundances of *Subdoligranulum variable* and *Dorea*, and lower relative abundances of *Eubacterium ventriosum* and *Ruminococcus torques*.¹¹ In a study using oral administration of encapsulated faeces for rCDI, faecal samples of patients before FMT had low diversity and relative abundances of Firmicutes and Bacteroidetes, and a high relative abundance of members of the Proteobacteria, specifically Enterobacteriaceae. Following FMT, subjects who developed recurrence of CDI had greater relative abundances of Enterobacteriaceae and did not show resolution of dysbiosis.⁵⁷ While it is not immediately clear how resident microbial communities in recipients influence FMT outcome, since diverse microbial taxa have been associated with unsuccessful FMT, it is postulated that they form a 'barrier' against uptake of donor microbial populations by creating competition for ecological niches and/or resources.

Pretreating recipients with antibiotics before FMT

Not all studies adopt the use of antibiotics prior to FMT and data remain controversial. Consistent with concept of species competition resulting in reduced FMT success, use of antibiotics prior to FMT may help alter resident gut microbiota in recipients to increase FMT efficacy. This is supported by a meta-analysis of

human studies and two studies in mice showing that pretreatment with antibiotics may enhance FMT efficacy.^{58–60} The inference based on these data is that antibiotics eliminate competition from resident micro-organisms by freeing previously occupied 'niches'. For instance, combination therapy of FMT and antibiotics was more effective than antibiotic cocktails or FMT therapy alone in restoring Bacteroidetes diversity in UC.⁶¹ In another study, patients with UC received FMT capsules containing purified Firmicutes spores (SER-287) with killed bacteria, parasites, fungi and viruses. A subset of recipients received vancomycin pre-FMT, which resulted in higher rates of clinical remission compared with recipients without vancomycin,^{62,63} providing further support for the 'ecological niche' hypothesis. Future studies should specifically assess the role of antibiotics prior to FMT in different conditions and its cost-effectiveness.

FORMULATING FMT PREPARATIONS AND DELIVERY PROCEDURES

New developments in FMT delivery

There is interest in developing convenient, easy-to-administer FMTs as current infusions are delivered via endoscopy (eg, colonoscopy or nasojejunal), enema or colonic transendoscopic enteral tubing (TET).^{64–66} Oral capsules has been developed showing efficacy comparable to delivery by colonoscopy regardless of whether fresh, frozen or lyophilised stools were used.^{67–69} However, frequency of doses and optimal overall duration is still unclear as study parameters were not directly comparable across

Figure 2 Potential mechanisms of faecal microbiota transplantation (FMT). Potential mechanisms of FMT include direct interaction or competition between donor and recipient gut microbiota to achieve homeostasis, effect of donor microbiota on the host immunity and effect of microbiota on modulating host metabolism and physiology. Competitive niche exclusion is a plausible mechanism behind the therapeutic effects of FMT in the treatment of *Clostridioides difficile* infections (CDI) by introducing competitors and/or modifying the diseased gut environment to the detriment of *Clostridioides difficile*. For example, introduction of non-toxigenic *C. difficile* strains can reduce the recurrence of CDI in subjects.¹ Another competition-based strategy harnessed by gut micro-organisms is the production of bacteriocins.² Quorum sensing systems have also been detected in *C. difficile* and are known to affect toxin expression profiles in other coexisting micro-organisms. FMT can alter host bile acid metabolism concomitant with alterations to gut microbiota composition. In CDI, a consistent metabolic signature with reduced primary bile acids and increased secondary bile acid production capacity has been reported after FMT. FMT restores Firmicutes phylum bacteria and secondary bile acid metabolism,³ providing a prime example of FMT re-establishing normal gut microbiota and host metabolism. FMT also has the ability to restore gut microbiota and mucosal immunity and systemic immunity of the host. In mice models of colitis, FMT reduced colonic inflammation and initiated a simultaneous activation of different immune pathways, leading to interleukin (IL-10) production by innate and adaptive immune cells including CD4+ T cells, invariant natural killer T (iNKT) cells and antigen-presenting cells (APC), and reduced the ability of macrophages, monocytes and dendritic cells to present MHCII-dependent bacterial antigens to colonic T cells.⁴ These results demonstrate the immunomodulating capability of FMT to therapeutically control intestinal homeostasis and highlight FMT as a valuable therapeutic option in immune-related pathologies. For instance, a reconstitution of the gut microbiome and a relative increase in the proportion of Foxp3+ regulatory T cells within the colonic mucosa were potential mechanisms through which FMT could abrogate immune check point inhibitors associated toxicity.⁵ online supplementary file 1 Please refer to Appendix 3 for references. iNKT, invariant natural killer T; MHCII, major histocompatibility complex class II.

different studies. So far, studies on oral capsule-based FMTs in chronic diseases have been limited to short-term outcomes (up to 8 weeks). The next steps must evaluate parameters such as dosage frequency and total treatment duration, as well as long-term response (years) of the recipient's gut microbiota.

Factors to consider in FMT formulations, preparations and delivery

Fresh, frozen or lyophilised (freeze-dried) stool

Figure 3 summarise FMT preparations, formulations and delivery methods. Although FMT infusions prepared from fresh or frozen stool may have comparable efficacy,^{70,71} various studies reported a decline in microbial viability in frozen stool. FMT oral capsules frozen up to 1 year were associated with a CDI resolution rate of 88%,⁶⁹ comparable to the widely reported 90% resolution rate for this condition. Quantitative PCR analyses showed statistically significant decreases in *Bifidobacterium* and *Lactobacillus* abundances in frozen and lyophilised FMT samples stored for 9 months.⁷² An RCT estimated rCDI remission rates associated with the use of fresh, frozen or lyophilised stools at 100%, 83%, and 78%, respectively, when delivered by colonoscopy.⁷³ In CD, contrasting results have been reported whereby one study reported increased clinical improvement and remission using fresh stool compared with frozen specimens,⁷⁴ while a meta-analysis found no significant difference in clinical

remission rates between fresh and frozen FMT preparations.⁷⁵ These observations raise the question about whether viability of micro-organisms in frozen stools is important for FMT success; in a pilot study of four patients, lyophilised sterile faecal filtrates and whole stool achieved similar efficacy for rCDI.⁷⁶

Aerobic or anaerobic preparations

Many bacterial populations in the human gut are obligate or facultative anaerobes, whose viability may depend on how FMT infusions are prepared. Comparisons conducted in rCDI subjects using aerobically and anaerobically prepared FMT infusions showed similar efficacy,⁷⁷ but comparisons have not been undertaken in other indications.⁷⁸ In a study of 73 active UC, treatment with 1 week of anaerobically prepared donor FMT resulted in higher steroid-free remission rates at 8 weeks compared with autologous FMT (32% vs 9%).⁶ The role of anaerobic FMT preparation in preserving anaerobes is consistent with the peptide nucleic acid sequencing analyses which showed that anaerobic preparation better preserves obligate anaerobes such as *Faecalibacterium prausnitzii*, an anti-inflammatory commensal bacterium whose absence is correlated with active IBD.⁷⁹

FMT dose amount and frequency

For traditional FMT delivery via enema and colonoscopy, most studies report the use of approximately 50 g of donor stool. The

Figure 3 For faecal microbiota transplantation (FMT) administration, lower GI delivery can be achieved through enema or colonoscopy, whereas upper GI delivery is through infusion via gastroscopy or via nasoenteric tubes. Oral capsules are non-invasive and is an ideal delivery method for FMT. Formulation to improve delivery, ensuring organism survival and ensure colonisation along the GI tract are needed.⁶ An encapsulation technique based on a water-in-oil emulsion can provide better insulation of FMT material from GI environment.⁷ One potential solution is the use of bile acid binding agents such as cholestyramine, when combined with Vcaps HPMC capsules, provided a 1700-fold increase in freeze-dried and rehydrated *Lactobacillus casei* in a simulated intestinal fluid containing 1% bile.⁸ Alternatively, a layer-by-layer encapsulation approach using mucoadhesive polysaccharides, chitosan and alginate has been shown to improve cytoprotection, bioavailability and engraftment of *Bacillus coagulans* in porcine and human intestines.⁹ Capsule design content release is dependent on intestinal stimuli.¹⁰ Other approaches include genetic engineering, for example, *Escherichia coli* engineered to bind to colorectal cancer cell receptors and catalyse production of molecules with anticancer activity.¹¹ This novel approach can be considered for FMT if constituent strains that contribute to clinical efficacy are identified. They can then be engineered to improve binding and colonisation in the gut of FMT recipients. Please refer to Appendix 3 for references. (online supplementary file 1)

basis for this is unclear, but a systematic review reported that the amount used had little impact on rCDI resolution rates. However, when less than 50 g of stool was administered, the relapse rate was fourfold higher (4% compared with 1%).⁸⁰ A recent meta-analysis also confirmed that faecal amount ≤ 50 g was associated with lower efficacy rates after a single infusion in rCDI.⁸¹ In IBD, approximately 50 cm³ faecal microbiota after filtration and centrifugation from stool was used. To determine the dosage required for oral FMT capsules, a comparative cohort analysis comparing clinical cure rates between 22.5 and 45 g of stool found little difference.⁸² No studies have investigated direct comparison of different FMT dosing for IBD; however, a study has evaluated enriching microbiota from the stool for the treatment of IBD.⁶⁵ Generally there is limited information to guide current practice related to FMT stool dose and frequency of administration.¹³ Stool mass varies among donors.⁴³ A more objective method to standardise FMT dosage such as microbial richness and diversity remains to be tested.

FMT delivery routes

Traditional/endoscopic FMT delivery

Traditional delivery routes of FMT include enema or colonoscopy, or gastroscopy or nasoenteric tubes.⁸³ Colonic TET has been used in specific centre in China.⁸⁴ In delivery through the lower GI tract, there is a trade-off between reliable delivery (ie, stool retention)

and invasiveness of the procedure. Delivery through enemas, colonoscopy, gastroscopy and nasoenteric tubes have comparable efficacy.⁷¹ Aspiration pneumonia and regurgitation are potential adverse events associated with gastric route FMT.

Oral capsules

In an RCT, the performance of FMT delivered via oral capsules containing frozen faecal material was equal to delivery by colonoscopy in rCDI.⁶⁷ Oral FMT capsules are prepared by mixing stools with a cryoprotectant (eg, glycerol), followed by double or triple encapsulation to protect the FMT material from gastric milieu. Capsule shells can be selected depending on the intended released site, with commercially available shells usually targeting gastric (FMTgr) or colonic release (FMTcr). A comparison between FMTgr and FMTcr capsules reported improved remission rates for treatment of rCDI using colonic release capsules (81% vs 75%), presumably due to the lower exposure of FMT material to chemical and enzymatic degradation.⁸² Various modifications can also be made to the capsule contents, such as used frozen oral capsules containing spores isolated from stool of healthy donors resulting in a 97% rCDI cure rate.⁸⁵ Another trial that used lyophilised faecal matter in oral capsules reported the same remission rate as enema delivery in rCDI subjects,⁸⁶ although there was reduced acquisition of *Bacteroidia* and *Verrucomicrobia* in patients treated with lyophilised oral capsules. RBX7455, a lyophilised non-frozen room temperature oral FMT capsule reported 90% primary treatment efficacy for rCDI.⁸⁷ Similarly, CP101, also based on lyophilised faecal matter, was as effective in low dose as high doses of 24–27 capsules, achieving 88% clinical success for rCDI treatment.⁶⁹ In another double-blind, placebo-controlled trial, 58 UC subjects were randomised to receive an 8-week treatment of either SER-287 (a mix of spores from members of the Firmicutes phylum) or placebo. Patients who achieved clinical remission after treatment with SER-287 exhibited decreased expression of UC-related genes and increased expression of non-IBD genes.⁸⁸

CONCLUSION

Considerable effort is being put into identifying factors that contribute to FMT success. While the vast majority of FMT studies are related to treatment of rCDI, a growing body of clinical evidence showed FMT efficacy in non-rCDI indications. Unlike rCDI in which there is a clear gut microbiota dysbiosis, the aetiology of other chronic conditions, such as IBD, IBS and obesity show stronger genetic and environmental predispositions. FMT success may depend more heavily on the capability of the donor microbiota to restore specific metabolic disturbances associated with each disease. Therefore, for FMT to become more effective, its delivery methods and microbial strains need to be rationally designed and tightly controlled taking into account important factors contributing to clinical success (eg, bacterial abundances, taxonomic composition, pretreatment with antibiotics, etc). Further characterisation of super-donors will result in the development of more refined FMT formulations to help standardise therapy and reduce variability in patient responses. In parallel, continued optimisation of FMT protocols, including a shift toward capsule-based delivery will help combat longevity issues associated with FMT and create a more patient-friendly alternative to current disease management strategies.

Author affiliations

¹Center for Gut Microbiota Research, The Chinese University of Hong Kong, Hong Kong, China

²Department of Medicine and Therapeutics, Institute of Digestive Disease, State Key Laboratory of Digestive Disease, LKS Institute of Health Sciences, The Chinese University of Hong Kong, Hong Kong, China

³St Vincent's Hospital and University of Melbourne, Melbourne, Victoria, Australia

⁴Department of Microbiology and LKS Institute of Health Sciences, The Chinese University of Hong Kong, Hong Kong, China

⁵Gastroenterology, Dayanand Medical College and Hospital, Ludhiana, India

⁶Department of Medicine, Shiga University of Medical Science, Otsu, Japan

⁷Department of Gastroenterology, Fujita Health University School of Medicine, Toyoake, Japan

⁸Department of Gastroenterology and Hepatology, Guangzhou Digestive Disease Center, Guangzhou First People's Hospital, School of Medicine, South China University of Technology, Guangzhou, China

⁹Gleneagles Medical Centre and Duke-NUS Medical School, Singapore, Singapore

¹⁰Department of Medicine, Faculty of Medicine, Chulalongkorn University and King Chulalongkorn Memorial Hospital, Bangkok, Thailand

¹¹National Gastric Cancer and Gastrointestinal Diseases Research Center, Pathumthani, Thailand

¹²Graduate Institute of Clinical Medical Science, China Medical University, Taichung, Taiwan

¹³Division of Gastroenterology, Taipei Veterans General Hospital, Taipei, Taiwan

¹⁴Medical Center for Digestive Diseases, The Second Affiliated Hospital of Nanjing Medical University, Nanjing, China

¹⁵Division of Microbiopathy, Sir Run Run Shaw Hospital, Nanjing Medical University, Nanjing, China

¹⁶Department of Medicine, Jichi Medical University, Shimotsuke, Tochigi, Japan

Contributors SCN, MAK, FKLC planned the meeting and established the main topics. WT and TZ performed the systematic search. All panel members were involved in presenting the literature in a Frontiers meeting in Hong Kong in June 2018. All panel members provided supporting evidence and drafted the text of discussion relevant to their topic. SCN, TZ and YKY wrote the initial draft of the manuscript. All panel faculties read and revised the manuscript for important intellectual content and approved the final manuscript. CJO, VM and KS are representatives of APAGE and NO is representative of APSDE.

Funding Funding from the Center for Gut Microbiota Research, Faculty of Medicine, The Chinese University of Hong Kong to support the Frontiers Meeting in Hong Kong, and Martin Ho (Imperial College London) for his input in the section on new deliveries of FMT.

Competing interests FZ invented the concept of GenFMTer and TET and devices related to it.

Patient consent for publication Not required.

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement All data relevant to the study are included in the article or uploaded as supplementary information.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>.

ORCID iDs

Siew C Ng <http://orcid.org/0000-0002-6850-4454>

Yun Kit Yeoh <http://orcid.org/0000-0003-0241-6117>

Francis K L Chan <http://orcid.org/0000-0001-7388-2436>

REFERENCES

- Mullish BH, Quraishi MN, Segal JP, *et al.* The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines. *Gut* 2018;67:1920–41.
- Krajicek E, Fischer M, Allegretti JR, *et al.* Nuts and bolts of fecal microbiota transplantation. *Clin Gastroenterol Hepatol* 2019;17:345–52.
- Moayyedi P, Surette MG, Kim PT, *et al.* Fecal microbiota transplantation induces remission in patients with active ulcerative colitis in a randomized controlled trial. *Gastroenterology* 2015;149:102–9.
- Rossen NG, Fuentes S, van der Spek MJ, *et al.* Findings from a randomized controlled trial of fecal transplantation for patients with ulcerative colitis. *Gastroenterology* 2015;149:110–8.
- Paramsothy S, Kamm MA, Kaakoush NO, *et al.* Multidonor intensive faecal microbiota transplantation for active ulcerative colitis: a randomised placebo-controlled trial. *The Lancet* 2017;389:1218–28.
- Costello SP, Hughes PA, Waters O, *et al.* Effect of fecal microbiota transplantation on 8-Week remission in patients with ulcerative colitis. *JAMA* 2019;321:156–64.
- Xu D, Chen VL, Steiner CA, *et al.* Efficacy of fecal microbiota transplantation in irritable bowel syndrome: a systematic review and meta-analysis. *Am J Gastroenterol* 2019.
- Bajaj JS, Kassam Z, Fagan A, *et al.* Fecal microbiota transplant from a rational stool donor improves hepatic encephalopathy: a randomized clinical trial. *Hepatology* 2017;66:1727–38.
- Bajaj JS, Fagan A, Gavis EA, *et al.* Long-Term outcomes of fecal microbiota transplantation in patients with cirrhosis. *Gastroenterology* 2019;156:1921–3.
- Kang D-W, Adams JB, Coleman DM, *et al.* Long-Term benefit of microbiota transfer therapy on autism symptoms and gut microbiota. *Sci Rep* 2019;9:5821.
- Kootte RS, Levin E, Salojärvi J, *et al.* Improvement of insulin sensitivity after lean donor feces in metabolic syndrome is driven by baseline intestinal microbiota composition. *Cell Metab* 2017;26:611–9.
- Vrieze A, Van Nood E, Holleman F, *et al.* Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology* 2012;143:913–6.
- Lai CY, Sung J, Cheng F, *et al.* Systematic review with meta-analysis: review of donor features, procedures and outcomes in 168 clinical studies of faecal microbiota transplantation. *Aliment Pharmacol Ther* 2019;49:354–63.
- ClinicalTrials.gov 2019.
- Wilson BC, Vatanen T, Cutfield WS, *et al.* The Super-Donor phenomenon in fecal microbiota transplantation. *Front Cell Infect Microbiol* 2019;9:2.
- Cammarota G, Ianiro G, Tilg H, *et al.* European consensus conference on faecal microbiota transplantation in clinical practice. *Gut* 2017;66:569–80.
- Hoffmann D, Palumbo F, Ravel J, *et al.* Improving regulation of microbiota transplants. *Science* 2017;358:1390–1.
- Verbeke F, Janssens Y, Wynendaele E, *et al.* Faecal microbiota transplantation: a regulatory hurdle? *BMC Gastroenterol* 2017;17:128.
- Barnes D, Ng K, Smits S, *et al.* Competitively selected donor fecal microbiota transplantation: butyrate concentration and diversity as measures of donor quality. *J Pediatr Gastroenterol Nutr* 2018;67:185–7.
- Ott SJ, Waletzki GH, Rehman A, *et al.* Efficacy of sterile fecal filtrate transfer for treating patients with *Clostridium difficile* infection. *Gastroenterology* 2017;152:799–811.
- Broecker F, Russo G, Klumpp J, *et al.* Stable core virome despite variable microbiome after fecal transfer. *Gut Microbes* 2017;8:214–20.
- Zuo T, Wong SH, Lam K, *et al.* Bacteriophage transfer during faecal microbiota transplantation in *Clostridium difficile* infection is associated with treatment outcome. *Gut* 2018;67:634–43.
- Park H, Millan BT, Hotte N, *et al.* A11 effect of fecal microbial transplant on microbial and phage composition in patients with *Clostridium difficile* infection. *J Can Assoc Gastroenterol* 2018;1(suppl_1):20–1.
- Hoffmann C, Dollive S, Grunberg S, *et al.* Archaea and fungi of the human gut microbiome: correlations with diet and bacterial residents. *PLoS One* 2013;8:e66019.
- Schoch CL, Seifert KA, Huhndorf S, *et al.* Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for fungi. *Proc Natl Acad Sci U S A* 2012;109:6241–6.
- Zuo T, Wong SH, Cheung CP, *et al.* Gut fungal dysbiosis correlates with reduced efficacy of fecal microbiota transplantation in *Clostridium difficile* infection. *Nat Commun* 2018;9:3663.
- Erb Downward JR, Falkowski NR, Mason KL, *et al.* Modulation of post-antibiotic bacterial community reassembly and host response by *Candida albicans*. *Sci Rep* 2013;3:2191.
- Reyes A, Wu M, McNulty NP, *et al.* Gnotobiotic mouse model of phage-bacterial host dynamics in the human gut. *Proc Natl Acad Sci U S A* 2013;110:20236–41.
- Wheeler ML, Limon JJ, Bar AS, *et al.* Immunological consequences of intestinal fungal dysbiosis. *Cell Host Microbe* 2016;19:865–73.
- Vermeire S, Joossens M, Verbeke K, *et al.* Donor species richness determines faecal microbiota transplantation success in inflammatory bowel disease. *ECCOIC* 2016;10:387–94.
- Kump P, Wurm P, Gröchenig HP, *et al.* The taxonomic composition of the donor intestinal microbiota is a major factor influencing the efficacy of faecal microbiota transplantation in therapy refractory ulcerative colitis. *Aliment Pharmacol Ther* 2018;47:67–77.
- Paramsothy S, Nielsen S, Kamm MA, *et al.* Specific bacteria and metabolites associated with response to fecal microbiota transplantation in patients with ulcerative colitis. *Gastroenterology* 2019;156:1440–54.
- Fuentes S, Rossen NG, van der Spek MJ, *et al.* Microbial shifts and signatures of long-term remission in ulcerative colitis after faecal microbiota transplantation. *Isme J* 2017;11:1877–89.
- Kazerouni A, Wein LM. Exploring the efficacy of pooled stools in fecal microbiota transplantation for Microbiota-Associated chronic diseases. *PLoS One* 2017;12:e0163956.
- Conceição-Neto N, Deboutte W, Dierckx T, *et al.* DOP080 low viral richness at baseline in ulcerative colitis associated with faecal microbiota transplantation success. *Journal of Crohns Colitis* 2017;11(suppl_1):S73–S74.
- Smillie CS, Sauk J, Gevers D, *et al.* Strain tracking reveals the determinants of bacterial engraftment in the human gut following fecal microbiota transplantation. *Cell Host Microbe* 2018;23:229–40.

- 37 SS L, Zhu A, Benes V, *et al.* Durable coexistence of donor and recipient strains after fecal microbiota transplantation. *Science* 2016;352:586–9.
- 38 Liu Q, Yang K, Xu Z, *et al.* 625 – donor fungi and bacteria engraftment after successful eradication of carbapenem-resistant Enterobacteriaceae with fecal microbiota transplantation: serial fecal Metagenomics analysis. *Gastroenterology* 2019;156:S-131.
- 39 Zhang F, Yeoh YK, Cheng F, *et al.* Tu1879 – rapid and durable engraftment of donor fungi and bacteria after successful fecal microbiota transplantation in acute graft-versus-host disease: intensive serial Metagenomics study. *Gastroenterology* 2019;156:S-1157–0.
- 40 Yeoh YK, Chen Z, Hui M, *et al.* Impact of inter- and intra-individual variation, sample storage and sampling fraction on human stool microbial community profiles. *PeerJ* 2019;7:e6172.
- 41 Seekatz AM, Aas J, Gessert CE, *et al.* Recovery of the gut microbiome following fecal microbiota transplantation. *MBio* 2014;5:e00893–14.
- 42 Fuentes S, van Nood E, Tims S, *et al.* Reset of a critically disturbed microbial ecosystem: faecal transplant in recurrent *Clostridium difficile* infection. *Isme J* 2014;8:1621–33.
- 43 Khanna S, Vazquez-Baeza Y, González A, *et al.* Changes in microbial ecology after fecal microbiota transplantation for recurrent *C. difficile* infection affected by underlying inflammatory bowel disease. *Microbiome* 2017;5.
- 44 Joseph OD, Thompson SV, Bogner A, *et al.* Longitudinal study of the human gastrointestinal microbiota following fecal microbiota transplant (FMT) for *Clostridium difficile* infections. *Faseb J* 2017;31.
- 45 Rothschild D, Weissbrod O, Barkan E, *et al.* Environment dominates over host genetics in shaping human gut microbiota. *Nature* 2018;555:210–5.
- 46 Lozupone CA, Stombaugh JJ, Gordon JL, *et al.* Diversity, stability and resilience of the human gut microbiota. *Nature* 2012;489:220–30.10.1038/nature11550
- 47 Tatusov RL, Galperin MY, Natale DA. The COG database: a tool for genome-scale analysis of protein functions and evolution. *Nucleic Acids Res* 2000;28:33–6.
- 48 Kanehisa M, Sato Y, Kawashima M, *et al.* Kegg as a reference resource for gene and protein annotation. *Nucleic Acids Res* 2016;44:D457–D462.
- 49 Almeida A, Mitchell AL, Boland M, *et al.* A new genomic blueprint of the human gut microbiota. *Nature* 2019;568:499–504.
- 50 Pasoli E, Asnicar F, Manara S, *et al.* Extensive unexplored human microbiome diversity revealed by over 150,000 genomes from metagenomes spanning age, geography, and lifestyle. *Cell* 2019;176:649–62.
- 51 Ianiro G, Valerio L, Masucci L, *et al.* Predictors of failure after single faecal microbiota transplantation in patients with recurrent *Clostridium difficile* infection: results from a 3-year, single-centre cohort study. *Clinical Microbiology and Infection* 2017;23:337.e1–337.e3.
- 52 Fischer M, Kao D, Mehta SR, *et al.* Predictors of early failure after fecal microbiota transplantation for the therapy of *Clostridium difficile* infection: a multicenter study. *Am J Gastroenterol* 2016;111:1024–31.
- 53 Duvallet C, Gibbons SM, Gurry T, *et al.* Meta-Analysis of gut microbiome studies identifies disease-specific and shared responses. *Nat Commun* 2017;8.
- 54 Narula N, Kassam Z, Yuan Y, *et al.* Systematic review and meta-analysis: fecal microbiota transplantation for treatment of active ulcerative colitis. *Inflamm Bowel Dis* 2017;23:1702–9.
- 55 Petrof EO, Gloor GB, Vanner SJ, *et al.* Stool substitute transplant therapy for the eradication of *Clostridium difficile* infection: 'RePOOPulating' the gut. *Microbiome* 2013;1.
- 56 Emanuelsson F, Claesson BEB, Ljungström L, *et al.* Faecal microbiota transplantation and bacteriotherapy for recurrent *Clostridium difficile* infection: a retrospective evaluation of 31 patients. *Scand J Infect Dis* 2014;46:89–97.
- 57 Staley C, Vaughn BP, Graiziger CT, *et al.* Community dynamics drive punctuated engraftment of the fecal microbiome following transplantation using freeze-dried, encapsulated fecal microbiota. *Gut Microbes* 2017;8:276–88.
- 58 Keshтели AH, Millan B, Madsen KL. Pretreatment with antibiotics may enhance the efficacy of fecal microbiota transplantation in ulcerative colitis: a meta-analysis. *Mucosal Immunol* 2017;10:565–6.
- 59 Ji SK, Yan H, Jiang T, *et al.* Preparing the gut with antibiotics enhances gut microbiota reprogramming efficiency by promoting Xenomicrobiota colonization. *Front Microbiol* 2017;8:1208.
- 60 Millan B, Park H, Hotte N, *et al.* Antibiotics and bowel preparation enhance the ability of fecal microbial transplantation to reshape the gut microbiota in IL-10^{-/-} mice. Canadian Journal of Gastroenterology and Hepatology Conference 2016.
- 61 Ishikawa D, Takahashi M, Haga K, *et al.* P074 microbial composition is effectively improved by combination therapy with fecal microbial transplantation and multiple antibiotics for ulcerative colitis. *Inflamm Bowel Dis* 2018;24(suppl_1):S27–S.
- 62 Bharat M, Curran J, Herfarth HH, *et al.* 85 - SER-287, an Investigational Microbiome Therapeutic, Induces Remission and Endoscopic Improvement in a Placebo-Controlled, Double-Blind Randomized Trial in Patients with Active Mild-to-Moderate Ulcerative Colitis. *Gastroenterology* 2018;154:S-25–20.
- 63 Simmons S, Diao L, O'Brien E, *et al.* Tu2019 - Engraftment of Ser-287, an Investigational Microbiome Therapeutic, is Related to Clinical Remission in a Placebo-Controlled, Double-Blind Randomized Trial (Seres-101) in Patients with Active Mild to Moderate Ulcerative Colitis (UC). *Gastroenterology* 2018;154:S-1371–0.
- 64 Peng Z, Xiang J, He Z, *et al.* Colonic transendoscopic enteral tubing: a novel way of transplanting fecal microbiota. *Endoscopy International Open* 2016;04:E610–E613.
- 65 Ding X, Li Q, Li P, *et al.* Long-Term safety and efficacy of fecal microbiota transplant in active ulcerative colitis. *Drug Saf* 2019;42:869–80.
- 66 Wang JW, Wang YK, Zhang F, *et al.* Initial experience of fecal microbiota transplantation in gastrointestinal disease: a case series. *Kaohsiung J Med Sci* 2019.
- 67 Kao D, Roach B, Silva M, *et al.* Effect of Oral Capsule- vs Colonoscopy-Delivered Fecal Microbiota Transplantation on Recurrent *Clostridium difficile* Infection. *JAMA* 2017;318:1985–93.
- 68 Youngster I, Mahabamunage J, Systrom HK, *et al.* Oral, frozen fecal microbiota transplant (FMT) capsules for recurrent *Clostridium difficile* infection. *BMC Med* 2016;14:134.
- 69 Staley C, Hamilton MJ, Vaughn BP, *et al.* Successful resolution of recurrent *Clostridium difficile* infection using freeze-dried, encapsulated fecal microbiota; pragmatic cohort study. *American Journal of Gastroenterology* 2017;112:940–7.
- 70 Lee CH, Steiner T, Petrof EO, *et al.* Frozen vs Fresh Fecal Microbiota Transplantation and Clinical Resolution of Diarrhea in Patients With Recurrent *Clostridium difficile* Infection. *JAMA* 2016;315:142–9.
- 71 Ramai D, Zakhia K, Ofosu A, *et al.* Fecal microbiota transplantation: donor relation, fresh or frozen, delivery methods, cost-effectiveness. *Ann Gastroenterol* 2019;32:30–8.
- 72 Jiang Z-D, Alexander A, Ke S, *et al.* Stability and efficacy of frozen and lyophilized fecal microbiota transplant (FMT) product in a mouse model of *Clostridium difficile* infection (CDI). *Anaerobe* 2017;48:110–4.
- 73 Jiang ZD, Ajami NJ, Petrosino JF, *et al.* Randomised clinical trial: faecal microbiota transplantation for recurrent *Clostridium difficile* infection - fresh, or frozen, or lyophilised microbiota from a small pool of healthy donors delivered by colonoscopy. *Aliment Pharmacol Ther* 2017;45:899–908.
- 74 Cui B, Feng Q, Wang H, *et al.* Fecal microbiota transplantation through mid-gut for refractory Crohn's disease: safety, feasibility, and efficacy trial results. *J Gastroenterol Hepatol* 2015;30:51–8.
- 75 Fang H, Fu L, Wang J. Protocol for fecal microbiota transplantation in inflammatory bowel disease: a systematic review and meta-analysis. *Biomed Res Int* 2018;2018:8941340–11.
- 76 Kao DH, Roach B, Walter J, *et al.* A51 effect of lyophilized sterile fecal filtrate vs lyophilized donor stool on recurrent *Clostridium difficile* infection (RCDI): preliminary results from a randomized, double-blind pilot study. *J Can Assoc Gastroenterol* 2019;2(Supplement_2):101–2.
- 77 Khanna S. Microbiota replacement therapies: innovation in gastrointestinal care. *Clin Pharmacol Ther* 2018;103:102–11.
- 78 Kelly CR, Khoruts A, Staley C, *et al.* Effect of Fecal Microbiota Transplantation on Recurrence in Multiply Recurrent *Clostridium difficile* Infection. *Ann Intern Med* 2016;165:609–16.
- 79 Chu ND, Smith MB, Perrotta AR, *et al.* Profiling living bacteria informs preparation of fecal microbiota transplantations. *PLoS One* 2017;12:e0170922.
- 80 Gough E, Shaikh H, Manges AR. Systematic review of intestinal microbiota transplantation (fecal bacteriotherapy) for recurrent *Clostridium difficile* infection. *Clin Infect Dis* 2011;53:994–1002.
- 81 Ianiro G, Maida M, Burisch J, *et al.* Efficacy of different faecal microbiota transplantation protocols for *Clostridium difficile* infection: A systematic review and meta-analysis. *United European Gastroenterol J* 2018;6:1232–44.
- 82 Allegretti JR, Fischer M, Sagi SV, *et al.* Fecal microbiota transplantation capsules with targeted colonic versus gastric delivery in recurrent *Clostridium difficile* infection: a comparative cohort analysis of high and low dose. *Dig Dis Sci* 2019;64:1672–8.
- 83 Jørgensen SMD, Hansen MM, Erikstrup C, *et al.* Faecal microbiota transplantation: establishment of a clinical application framework. *Eur J Gastroenterol Hepatol* 2017;29:e36–45.
- 84 Zhang F, Zhang T, Zhu H, *et al.* Evolution of fecal microbiota transplantation in methodology and ethical issues. *Curr Opin Pharmacol* 2019;49:11–16.10.1016/j.coph.2019.04.004
- 85 Khanna S, Pardi DS, Kelly CR, *et al.* A Novel Microbiome Therapeutic Increases Gut Microbial Diversity and Prevents Recurrent *Clostridium difficile* Infection. *J Infect Dis* 2016;214:173–81.
- 86 Jiang Z-D, Jenq RR, Ajami NJ, *et al.* Safety and preliminary efficacy of orally administered lyophilized fecal microbiota product compared with frozen product given by enema for recurrent *Clostridium difficile* infection: a randomized clinical trial. *PLoS One* 2018;13:e0205064.
- 87 Blount K, Gerding DN, Khanna S, *et al.* 624 – comparison of two open-label trials demonstrates similar 6-month outcomes for Rbx2660 and Rbx7455—Investigational microbiota restoration therapeutics administered by enema or oral capsules for preventing recurrent *Clostridium difficile* infections. *Gastroenterology* 2019;156:S-130–S-131.
- 88 Diao L, Nnamani MC, OBrien E, *et al.* 623 – Ser-287, an investigational microbiome therapeutic, induces widespread transcriptional changes related to clinical remission in a placebo-controlled, double-blind randomized trial (Seres-101) in patients with active mild-to-moderate ulcerative colitis. *Gastroenterology* 2019;156:S-130.