Original research

Stool microRNA profiles reflect different dietary and gut microbiome patterns in healthy individuals

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ABSTRACT

Objectives MicroRNA (miRNA) profiles have been evaluated in several biospecimens in relation to common diseases for which diet may have a considerable impact. We aimed at characterising how specific diets are associated with the miRNome in stool of vegans, vegetarians and omnivores and how this is reflected in the gut microbiological composition, as this is still poorly explored.

Design We performed small RNA and shotgun metagenomic sequencing in faecal samples and dietary recording from 120 healthy volunteers, equally distributed for the different diets and matched for age.

Results We found 49 miRNAs differentially expressed among vegans, vegetarians and omnivores (adj. p <0.05) and confirmed trends of expression levels of such miRNAs in vegans and vegetarians compared with an independent cohort of 45 omnivores. Two miRNAs related to lipid metabolism, miR-636 and miR-4739, were inversely correlated to the non-omnivorous diet duration, independently of subject age. Seventeen miRNAs correlated (r(ho)>0.22, adj. p <0.05) with the estimated intake of nutrients, particularly animal proteins, phosphorus and, interestingly, lipids. In omnivores, higher Prevotella and Roseburia and lower Bacteroides abundances than in vegans and vegetarians were observed. Lipid metabolism-related miR-425-3p and miR-638 expression levels were associated with increased abundances of microbial species, such as Roseburia sp. CAG 182 and Akkermansia muciniphila, specific of different diets. An integrated analysis identified 25 miRNAs, 25 taxa and 7 dietary nutrients that clearly discriminated (area under the receiver operating characteristic curve=0.89) the three diets.

Conclusion Stool miRNA profiles are associated with specific diets and support the role of lipids as a driver of epigenetic changes and host-microbial molecular interactions in the gut.

INTRODUCTION

Dietary habits are strongly related to lifestyle factors as well as social life and culture, and all are strictly connected with health. Diet and nutrition represent relevant risk factors for different metabolic and chronic diseases such as steatosis and non-alcoholic fatty liver disease, atherosclerosis and cancer.1

Significance of this study

What is already known on this subject?

⇒ The adoption of a vegetarian or vegan diet is associated with a lower intake of relevant risk factors for chronic diseases, such as cardiovascular diseases or cancer. However, it is still controversial whether such diets are always healthier and recommended for all.

⇒ Dietary components can modulate the composition and function of the gut microbiome and serve as a substrate for bacterial metabolism that could influence the host health.

⇒ A host-gut interplay may modulate the effects of the diet. MicroRNAs (miRNAs) found in stool may directly regulate specific bacterial gene expression and affect gut microbial growth, therefore being essential for maintaining normal gut microbiota.

What are the new findings?

⇒ Diet is associated with stool miRNome profile of omnivores, vegetarians and vegans, and several miRNAs characterise peculiar signatures of the different dietary regimes.

⇒ MiR-636 and miR-4739, having a significant role in lipid metabolism, show expression levels inversely related to the duration of the non-omnivorous dietary regime, independently of age. Furthermore, targets of stool miRNAs upregulated in vegetarian/vegan subjects are enriched in both lipid and folate metabolism.

⇒ Vegan and vegetarians are characterised by high stool levels of miR-425-3p, an miRNA related to lipid metabolism, whose levels in stool correlate with the relative abundance of specific microbial taxa, including Akkermansia muciniphila.

⇒ Stool miRNA profiles and microbiome composition, together with dietary nutrients, can accurately distinguish different diets and strengthen the evidence of host-gut microbiome crosstalk.
In monitoring adherence to specific dietary regimes, where the intake of nutrients should be carefully checked, a translational approach of a non-invasive stool miRNA profiling may be easily implemented.

An integrated analysis of gut microbiome and faecal miRNAs may provide insights for designing more accurate personalised dietary patterns and recommendations aimed at prevention/treatment of chronic diseases.

High-level consumption of red, processed meat and animal fats have been associated with an increase in cancer risk, including colorectal cancer (CRC), while the high intake of dietary fibres is considered a protective factor.

An increasing number of studies showed that vegetarian (VT) and vegan (VN) dietary patterns are associated with significantly lower levels of the most relevant risk factors for chronic diseases (such as body mass index (BMI) or lipid and glucose levels) with a consequent reduced risk of incidence or mortality for ischaemic heart disease or cancers. However, it is difficult to discern whether the health advantages attributed to VNs could be generalised to VTs or even to moderate meat eaters following a healthy diet. Moreover, there is still the need to elucidate the precise molecular mechanisms of these associations. So far, only very few studies have performed rigorous experiments comparing omnivorous, VT and VN subjects as distinct experimental groups. The few available, investigating the molecular mechanisms involved with dietary components absorption, were generated from cell cultures and animal models.

In the past few years, microRNAs (miRNAs), small non-coding RNAs that modulate gene expression at the post-transcriptional level, have been studied in detail as important binding molecules and, as a consequence, in all the miRNA-mediated molecular mechanisms. Based on an miRNA candidate approach, we performed a pilot study on stool/plasma samples of healthy volunteers with different dietary habits and found differential miRNA expression levels among them. Different studies reported a relationship between the expression of faecal miRNAs and gastrointestinal disorders, including CRC. Then, stool samples can represent a proper non-invasive surrogate tissue to evaluate the transcriptional and epigenomic changes occurring on the adherence to a different dietary regimen. However, many issues remain unsolved regarding the quality and quantity of dietary components needed to regulate miRNA expression in humans. Recently, we clarified certain relationships between specific miRNA expression and the inclusion of specific nutrients in the diets of healthy individuals.

Dietary components can modulate the composition and function of the gut microbiome and diets rich in fibres and poor in animal-based products (VN/VT regimes, Mediterranean diet) may drive a selection of specific taxa. In parallel, a growing number of studies reported the association of a gut microbiome dysbiosis with several gastrointestinal and non-gastrointestinal diseases, including diabetes, inflammatory bowel disease and cancer.

Liu and colleagues have demonstrated that faecal miRNAs directly regulate specific bacterial gene expression and affect gut microbial growth, being therefore essential for the maintenance of normal gut microbiota. Moreover, an integrated analysis of the gut microbiome and small non-coding RNAs in the human stool may provide insights for designing more accurate tools for diagnostic purposes in CRC. Intestinal microbiota and miRNAs may interact to regulate host gene expression. However, the interplay existing between gut microbiome and miRNA expression profiles in the presence of a specific dietary regime is still unknown.

In the present study, the relationship between miRNA profiles and the gut microbiome according to different long-term dietary regimes was explored. Small non-coding RNA sequencing was performed in faecal and plasma samples from 120 healthy volunteers with omnivorous, VT and VN dietary habits. In addition, gut microbiome was analysed by shotgun metagenomics. The results from sequencing were integrated with detailed information on food and specific nutrient intake by validated questionnaires to elucidate the existence of molecular characterisation of a specific dietary regime.

By DESeq2 analysis, we found 49 miRNAs differentially expressed in stool of VNs and VTs in comparison with omnivores (Os) (adj. p<0.05). Interestingly, differentially expressed miRNAs among the dietary groups are involved in the regulation of lipid and folate metabolism, adipocyte differentiation and connected with obesity and obesity-connected diabetes. A combination of these miRNAs, microbial taxa and nutrients clearly distinguished subjects according to their diet (area under the receiver operating characteristic curve (AUC)=0.89).

**METHODS**

Detailed methods are provided in the online supplemental materials of the manuscript.

**Study population recruitment and dietary information**

A cohort of 120 healthy individuals (72 women and 48 men), categorised as VTs, VNs and Os, was recruited on a voluntary basis in Turin (Italy) between May 2017 and July 2019. Eligible subjects were included in the study according to the following selection criteria: the associated dietary regime followed for more than 1 year, no consumption of antibiotics in the previous 3 months before sampling, no use of aspirin or other anti-inflammatory drugs in the month prior to the sampling, no evidence of intestinal and other pathologies, no pregnancy and lactation. Three subjects made use of antihypertensive drugs and only one made use of a cholesterol-lowering medication. The study cohort included the same proportion of VT, VN and O, matched for sex and age (figure 1A). Anthropometric data were collected for all subjects at the time of sampling. For an independent group of unmatched 45 omnivorous healthy individuals (31 women and 14 men), stool small RNA-Seq data (performed in the same laboratory and with the same protocol/pipeline of analysis) were also available for miRNA profiles validation.

All participants signed informed written consent. The design of the study, the informed consent and protocols were approved by the local Ethics Committee.

All recruited subjects filled in a validated self-administered Food Frequency Questionnaire (FFQ) assessing the usual diet, together with lifestyle and personal history data, in accordance with detailed questionnaires, validated food frequency questionnaires, and dietary assessment methods, as previously described in the study. Anthropometric data were also collected for all participants at the time of sampling, and detailed lifestyle and personal history data were collected for all participants at the time of sampling.
with the European Prospective Investigation into Cancer and Nutrition EPIC study standards.24 The FFQ consisted of 248 questions concerning 188 different food items and included photos with two or three sample dishes of definite sizes or references to standard portion sizes. The composition in nutrients of individual food items was obtained from Italian food composition tables25 and the average daily intake of macronutrients and micronutrients for each volunteer was estimated.

Collection of biological samples
Naturally evacuated faecal samples were obtained from all volunteers previously instructed to self-collect the specimen at home. Stool were collected in nucleic acid collection and transport tubes with RNA/DNA stabilising solution (Norgen Biotek Corp) and stored at −80°C until RNA and DNA extraction.27 Blood samples were collected into EDTA and serum clot activator tubes, according to standard phlebotomy procedures, when volunteers brought the stool samples to the laboratory. EDTA tubes were centrifuged at ×1000 g for 10 min at room temperature and plasma was aliquoted and stored at −80°C. Serum tubes were centrifuged at ×4000 g for 10 min at room temperature, aliquoted and stored at −80°C. One aliquot of serum was used for the analyses of vitamin B12 and ferritin. The time between blood sample procurement and plasma/serum storage was less than 3 hours.

Extraction of total RNA from stool and plasma
Total RNA was extracted from 200 µL of plasma with the miRNeasy plasma/serum mini kit (Qiagen, Hilden, Germany) using the QiaCube extractor (Qiagen) following the manufacturer’s instructions and as described in the study by Ferrero et al27 and Sabo et al.28 RNA concentration was quantified by Qubit microRNA Assay Kit (Invitrogen).

Extraction of total DNA from stool
The DNA extraction was performed with the QIAamp DNA stool MiniKit (QIAGEN) according to the instructions of the manufacturer and as described in the study by Tarallo et al12 and Thomas et al.22 The DNA quantification was performed with Qubit fluorometer (Qubit DNA HS Assay Kit; Invitrogen).

Small RNA sequencing and bioinformatic analyses
Library preparation for small RNA (sRNA) transcripts was performed with the NEBNext Multiplex Small RNA Library Prep Set for Illumina (Protocol E7330, New England BioLabs, USA) as described in the study by Ferrero et al.27 For each sample, 250 ng of RNA was used as starting material to prepare libraries.

The obtained sequence libraries were subjected to the Illumina sequencing pipeline and 50 cycles sequencing-by-synthesis on the HiSeq 2000 (Illumina, USA) (in collaboration with Genecore Facility at EMBL, Heidelberg, Germany). Small RNA sequencing analyses were performed using a previously described approach.12 The reads passing the quality control were trimmed from the adapter sequences using Cutadapt V1.10. Trimmed reads were mapped against human miRNA
sequences from miRBase V.22 \(^{29}\) using BWA V.0.7.12 \(^{30}\) with default settings. Summary statistics of the data and alignment are reported in online supplemental table 1A,B. Human miRNAs were annotated and quantified using two methods called the ‘knowledge-based’ and ‘position-based’ methods as described in Tarallo et al. \(^{12}\) miRNAs whose assigned arms were derived from the ‘position-based’ methodology were indicated in italic.

Differential expression analysis was performed with DESeq2 R package V.1.22.2 \(^{31}\) using the likelihood ratio test function including age and sex as covariates in the model. An miRNA was defined as differentially expressed (DEmiRNA) if associated with a Benjamini-Hochberg (BH)-adjusted p value <0.05 and supported by more than 15 reads in at least one of the dietary groups. The set of DEmiRNAs was overlapped with the annotation from Human microRNA Disease Database (HMDD) V.3.2, \(^{32}\) considering the annotations associated with each miRNA precursor and the database annotations related to gastrointestinal cancers, metabolic disorders, immune-related, or cardiovascular diseases.

**Shotgun metagenomic and bioinformatic analyses**

Libraries were prepared using the Nextera DNA Library Preparation kit (Illumina) and sequenced on an Illumina HiSeq platform as described in the study by Thomas et al. \(^{22}\)

Host contamination was removed using the human sequence removal procedure from the Human Microbiome Project. \(^{23}\) Raw reads were quality-trimmed (Phred score <25) and reads shorter than 60 bp were discarded with the SolexaQA++ software. \(^{34}\) The number of reads obtained for each sample is reported in online supplemental table 1C. Taxonomic profiling was carried out by using MetaPhlAn 3. \(^{35}\) Functional profiles were obtained by HUMAnN 3.0. \(^{36}\)

**Integration between miRNA expression, nutrient intake and taxa**

The R package mixOmics was employed for the integration of the three datasets (taxonomic profiles, miRNA expression profiles and dietary information), using the Data Integration Analysis for Biomarker discovery using Latent cOmponents (DIABLE) model. \(^{37}\) Datasets were integrated after normalisation (scale R function) and removal of near-zero variables (nearZeroVar function of R package caret).

**Statistical analyses**

All statistical analyses were performed using R software V.4.0.4. The analysis of the dietary and lifestyle covariates among subject groups was performed using the \(\chi^2\) test for categorical variables. For continuous covariates, Kruskal-Wallis and Wilcoxon rank-Sum tests were performed.

Correlation analysis between DEmiRNA expression and nutrient intake was performed using the Spearman method. Multiple testing correction was performed using the BH method and correlations associated with an adjusted p value <0.05 were considered as statistically significant.

The Generalized Linear Regression Model (GLM) was computed using the glm R function to assess the relevance of subject covariates in explaining the observed stool DEmiRNA levels. DEmiRNA levels were considered as the dependent variable of the model while subjects age, sex, BMI, physical activity, waist circumference and season of sample collection were considered as the independent variables of the model. A covariate was considered related to DEmiRNA expression if associated with a p value <0.05. The relationship between DEmiRNA expression and the estimated intake of nutrients was evaluated computing a GLM adjusted for age, sex and BMI. The significance of the GLM was computed with respect to a null model using F-test implemented in the ANOVA R function. Models associated with a p value <0.05 were considered statistically significant. The contribution of a nutrient to the model was considered significant if the associated p value was lower than 0.05 and lower than those contributed for the computation of sex, age and BMI to the model.

Classification accuracy of the features identified by the DIABLE analysis was performed using a 10-fold cross-validation approach with the Random Forest classifier implemented in Weka V.3.8.5 (https://www.cs.waikato.ac.nz/ml/weka/). The classification efficiency was evaluated in terms of AUC.

**MiRNA targets functional enrichment analysis**

Functional enrichment analysis was performed using RBiomirGS V.0.2.12 \(^{38}\) considering validated targets of: (1) DEmiRNAs coherently differentially expressed in VT and VN with respect to O; (2) miRNAs significantly correlated with cluster of nutrients or (3) taxa; (4) DEmiRNAs from the DIABLE analysis. The log2FC and adjusted p value computed between the VN and O diets were used as inputs for the tool. The gene sets characterised by an adjusted p value lower than 0.05 were considered as significantly enriched.

**MiRNA expression and epigenetic state analysis in intestinal tissues**

The analysis of miRNA expression in intestinal tissue samples was performed considering the data from the TissueAtlas, \(^{39}\) which includes miRNA expression levels measured in two colon and three small intestine samples. The epigenetic status of miRNA coding genes was evaluated using the chromatin status defined by the Roadmap Epigenomics Project. \(^{40}\) Specifically, the analysis was performed considering the 15 statuses (Core 15-state model) defined by integrating epigenetic data of duodenum, small intestine, colon, rectum mucosa, or colon smooth muscle samples. Only mature miRNAs with an univocal genomic locus were considered for the analysis.

**RESULTS**

**Study subjects and dietary characteristics**

A total of 120 volunteers, divided into 40 Os, 40 VTs and 40 VNs, were included in the study. The characteristics of the subjects and the design of the study are described in table 1 and figure 1A, respectively. Subjects following an omnivorous diet displayed a slightly higher BMI (kg/m\(^2\)) compared with the other groups (O: 23.6 ± 3.9, VT: 21.9 ± 2.6, VN: 21.8 ± 2.9 kg/m\(^2\), in all comparisons p < 0.05) and waist circumference (O: 83.8 ± 11.6, VT: 78.1 ± 8.6, VN: 78.9 ± 8.8 cm, in O vs VT, p < 0.05) (table 1). Days of sample collection were differently distributed for VT and VN with respect to O individuals with respect to the different seasons of the year (p < 0.05) (data not shown).

All the dietary and lifestyle characteristics are reported in detail in online supplemental table 2. In serum, the concentration of ferritin was significantly decreased in VT/VN with respect to O (p < 0.01 and p < 0.05, respectively), while vitamin B12 was lower in VT with respect to the other two groups. From the list of 188 items included in the EPIC questionnaires, the analysis was focused on the most representative nutrients characterising the differences in the dietary regimes (consumption of meat, processed meat, fish, cheese and dairy products, bread and pasta, total vegetables and fruits). The relative weekly intake...
of specific food and drinks as reported in the subject questionnaires is summarised in figure 1B and online supplemental figure 1A. A significantly higher intake of nutrients such as vitamin C, β-carotene and fibres was found in VN when compared with O. Conversely, in the same comparison, a significant decrease in vitamin D, animal proteins, retinol and cholesterol was observed in VN (figure 1C). In VT, the discrepancies with O were less marked but in line with those observed in VN: the only evident difference was in a decreased intake of polyunsaturated fatty acids in VT when compared with O (figure 1C). The distribution of the macronutrient and micronutrient intake among the three dietary groups is represented in online supplemental figure 1B.

### Stool miRNA profiles according to different diets

After sequencing of stool RNA samples, an average of 64,681 reads (0.83%) was aligned to known human miRNA sequences and a median number of 253, 321 and 310 stool miRNAs were detected in O, VT and VN, respectively (online supplemental table 1A). Among these miRNAs, 145 (59.43%) were represented in all three dietary groups (online supplemental table 1A). A significantly higher intake of nutrients such as vitamin C, β-carotene and fibres was found in VN when compared with O. Conversely, in the same comparison, a significant decrease in vitamin D, animal proteins, retinol and cholesterol was observed in VN (figure 1C). In VT, the discrepancies with O were less marked but in line with those observed in VN: the only evident difference was in a decreased intake of polyunsaturated fatty acids in VT when compared with O (figure 1C). The distribution of the macronutrient and micronutrient intake among the three dietary groups is represented in online supplemental figure 1B.

#### Table 1 Study population characteristics

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Omnivores</th>
<th>Vegetarians</th>
<th>P value</th>
<th>Vegans</th>
<th>P value</th>
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<tr>
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<tr>
<td></td>
<td>Males</td>
<td>16</td>
<td>16</td>
<td>16</td>
<td>16</td>
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<tr>
<td>Age (years)</td>
<td>Mean±SD</td>
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<td>40.6±11.7</td>
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<tr>
<td></td>
<td>Range</td>
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<td>21.1–67.4</td>
<td>18.5–60.6</td>
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<td>Years of vegetarian or vegan diet</td>
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<td>9.7±9.0</td>
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<td>4.8±2.5</td>
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<tr>
<td></td>
<td>Range</td>
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<td>BMI (kg/m²)</td>
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<td>78.9±8.8</td>
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<td>0.02</td>
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<td>14</td>
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<td></td>
<td>Current smoker</td>
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<td>2</td>
<td>4</td>
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</tr>
</tbody>
</table>

For continuous variables, the p value was computed using the Kruskal-Wallis method while χ² test was used for analysing categorical variables. Statistically significant values are in bold.

BMI, body mass index.

...
Twenty-two miRNAs were among those identified as differentially expressed in VT/VN compared with O (online supplemental table 4A). To further explore the relationship between the nutrient estimated intake and the stool DEmiRNA levels, a GLM regression analysis adjusted for sex, age and BMI was performed revealing 56 miRNA-nutrient associations supported by both analyses (figure 2E, online supplemental table 4B). The nutrients characterised by the highest number of related DEmiRNAs were plant lipids (eight miRNAs), followed by animal lipids (seven miRNAs), and total/animal proteins and phosphorus (six miRNAs). Conversely, among all DEmiRNAs, miR-8053 levels were related to the highest number of nutrients (n=9), followed by miR-4277 and miR-1260a-3p (for both n=8). Out of 56 miRNA-nutrient associations, 28 associations involving nutrients not specific to a dietary regime like animal proteins/lipids were coherent also even considering the three dietary groups separately.

The analysis of HMDD annotations showed that 12 out of the 49 stool DEmiRNAs were characterised by precursor sequences that were related to a specific disease phenotype in different studies (n=191) (online supplemental figure 3A and online supplemental table 3E).

**Validation in stool samples of an independent group of omnivores**

The expression profiles of the 49 stool DEmiRNAs were also investigated in an independent cohort of 45 O analysed in another project (see Methods section). All DEmiRNAs were detected in stool of the validation cohort and their levels strongly correlated with those of the 40 O investigated in this study (rho=0.82,
Conversely, in plasma samples, 11 xenomiRs were detected, (online supplemental table 6). For this xenomiR, no sequence was observed in the genome of other edible crops. The presence of food-derived non-human miRNAs (xenomiRs) was evaluated by analysing the fraction of human-unmapped reads. In stool samples, an average of 186 reads (0.005%) from the unmapped reads set was assigned to xenomiRs, while on plasma, the mapping rate was slightly higher (1290 reads on average, 0.87% of human-unmapped reads) (online supplemental table 1). Only two xenomiRs (gga-mir-1692 and gma-MIR6300) were detected above the expression threshold used in our analysis. Interestingly, the stool levels of gma-MIR6300, which is annotated in the soy genome (Glycine max), were significantly higher (p<0.05) in VT and VN with respect to O (online supplemental table 5C). For this xenomiR, no sequence similarity was found among human miRNAs, while a homologous sequence was observed in the genome of other edible vegetables and fruits (e.g., Solanum lycopersicum, Solanum tuberosum and Cucumis melo) (online supplemental table 6). Conversely, in plasma samples, 11 xenomiRs were detected, with chi-mir-30d (annotated to Capra hircus genome) characterised by the highest number of reads. All the 11 miRNAs were annotated to an animal species (C. hircus, Gallus gallus, Sus scrofa, Salmo salar or Gadus morhua). However, given the high homology of these miRNAs with their homologous sequence in humans, a real circulating level of these molecules is less reliable. The observed reads could be actually related to the sequencing of portions of the corresponding miRNA precursor in humans.

**Gut microbiota**

**Gut microbiome composition and functions and relation to faecal miRNAs**

Gut microbiome taxonomic composition varied according to the different dietary patterns. Among differently abundant taxa, Prevotella copri and Roseburia sp. CAG 182 showed higher abundance in VT/VN compared with O, while Bacteroides dorei (currently Phascolarctobacterium dorei) prevailed in O (p<0.05; figure 3A).

A functional profiling of the gut metagenomes was performed with HUMAnN3. Different gut microbiome composition is reflected in differential potential activities. The microbiome of O showed increased abundance of several metabolic pathways, including pathways related to the biosynthesis of amino acids, biogenic amines or their precursors (L-isoleucine biosynthesis IV, L-arginine biosynthesis I and II, super-pathway of L-lysine, L-threonine and L-methionine biosynthesis II, putrescine biosynthesis IV, chorismate biosynthesis from 3-dehydroquinicinate, as well to the biosynthesis of vitamins (formyl-tetrahydrofolate biosynthesis, pantothenate and coenzyme A biosynthesis III) (FDR<0.05; figure 3B).

Clustering analysis of the correlation coefficient between bacterial abundance and stool miRNA expression levels resulted in less homogeneous clusters (online supplemental figure 2C). Despite two main clusters of bacterial species and miRNAs were identified, correlation coefficients were significant only on specific bacteria-miRNA pairs. The bacterial species characterised by the highest number of correlated miRNAs (p<0.05) were Ruminococcaceae bacterium D5 (11 correlated miRNAs), P. copri (n=8) and Phascolarctobacterium succinatutens (n=7). Among the miRNAs correlated with bacterial species, 10 were significantly differentially expressed in VT/VN compared with O (online supplemental table 4A).

**Integration of miRNAs, microbiome and diet**

The DIABLO model identified a set of 25 DEmiRNAs, 25 taxa and 7 dietary nutrients able, all together, to discriminate the three groups (figure 3C). As for stool DEmiRNAs, a GLM adjusted for age, sex, BMI, physical activity, waist circumference and season of sample collection was applied to evaluate the relationship between taxa and subject covariates. Only five species were associated with a significant contribution of at least one of these variables (online supplemental table 3B). Partial least squares discriminant analysis and the hierarchical clustering showed a separation of subjects according to their diet (figure 3DE). To assess the robustness of the data reported in this study in discriminating the different dietary groups, a machine learning analysis using the combination of these features was applied. The analysis confirmed that the identified features were able to discriminate efficiently the different dietary groups (average AUC=0.89). As expected, higher accuracy was observed in classifying omnivorous (AUC=0.95) and VN (AUC=0.94) individuals than VT subjects (AUC=0.79).

Two DEmiRNAs, mir-425-3p and mir-638, attracted our interest since they were associated with a coherent higher expression in VT/VN compared with both the analysed O groups (among those not affected by BMI), they were also identified by DIABLO as discriminant features for the different dietary regimes, and they were significantly correlated with nutrients (figure 2E). Therefore, these DEmiRNAs were further investigated for their relationship with gut microbiome composition. Subjects within the highest quartile of miR-425-3p expression levels (regardless of the type of diet) showed a significantly higher abundance of Akkermansia muciniphila, Roseburia hominis, Roseburia sp. CAG 182, Oscillibacter sp. 57_20, Oscillibacter sp. 7a-3p, Ruminococcaceae bacterium D5 (11 correlated miRNAs), P. copri (n=8) and Phascolarctobacterium succinatutens (n=7). Among the miRNAs correlated with bacterial species, 10 were significantly differentially expressed in VT/VN compared with O (online supplemental table 4A).
Oscillibacter sp. CAG 241 and R. bacterium D5 compared with those in the lowest quartile (figure 3F and online supplemental figure 4), while Anaerotruncus colihominis and Flavonifractor plautii were more abundant in subjects with the lowest levels of the same miRNA. Consistently, higher levels of Roseburia sp. CAG 182 and R. bacterium D5, as well as a lower abundance of A. colihominis were found in subjects within the highest expression levels of miR-638.

Enrichment analysis

The miRNA target enrichment analysis was performed to explore whether independent sets of relevant stool miRNAs identified in this study (ie, all identified DEmiRNAs either upregulated or downregulated, miRNAs identified by DIABLO and miRNAs correlated with bacterial species or nutrient intake) were predicted to regulate common biological processes. A total of 927 functional terms were significantly enriched (adj. p<0.05) in target genes from each set of miRNAs considered (online supplemental table 7), with 68 terms enriched in at least three miRNA sets concomitantly (figure 4A). Out of them, 66 were enriched in miRNAs upregulated in VN/VT, suggesting an inhibition of the related process (negative coefficients in figure 4A). In particular, the terms that were redundantly enriched in the highest number of miRNA sets (five sets) were KEGG_One_carbon_pool_by_folate and KEGG_Glyoxylate_and_dicarboxylate_metabolism.
Gut microbiota

most represented in the analysis (figure 4B). These terms were enriched in the stool DEmiRNAs upregulated in VN/VT but also in miRNAs identified by DIABLO and those correlated with higher intake of nutrients observed in VN and VT. Finally, Arachidonic_acid_metabolism was the sole KEGG metabolic-related term enriched for DEmiRNAs downregulated in VN/VT.

Analysis of stool DEmiRNA expression and epigenetic state in intestinal tissue samples

Finally, we evaluated the expression levels of stool DEmiRNAs in intestinal tissues using the data from the TissueAtlas. The analysis confirmed that 19 DEmiRNAs out of 22 were detectable also in the intestine (online supplemental table 8A). In particular, miR-143-3p was the DEmiRNA characterised by the highest expression levels in colon samples (average Z-score=2.40), while others like miR-638, miR-425-3p and miR-636 were detected in colon/small intestine as well as in different tissue samples of the TissueAtlas repository.

Furthermore, since the epigenetic status of a gene locus can represent a reliable proxy of an active/repressive transcriptional state, the epigenetic state of DEmiRNA coding genes was evaluated in the Roadmap Epigenomics Project. The analysis confirmed a transcriptionally active epigenetic state in 41 out of the 49 stool DEmiRNAs in at least one of the sample types analysed (online supplemental table 8B). Specifically, colon mucosa was associated with the highest number of epigenetically active DEmiRNA loci (n=26), followed by duodenum mucosa (n=23), jejunum mucosa (n=23), ileum mucosa (n=23), and large intestine (n=23).
small intestine and colon smooth muscle tissues (n=18). The analysis confirmed, in colon smooth muscle samples, a chromatin state related to a strong transcription at miR-143 coding locus coherently with previous observations.41

**DISCUSSION**

In the present study, we explored the facetal miRNome of 120 healthy subjects adhering to a VN, VT or omnivorous diet, reporting that miRNA expression levels are affected by diet, with a possible relationship with the gut microbiota composition and function. Forty-nine miRNAs were significantly differentially expressed among the three dietary groups and 15 of them showed a consistent trend of expression of upregulation or downregulation in both VT and VN in comparison with two independent groups of O. These findings suggest a gradual adaptation of the miRNome in response to a specific dietary regimen, probably as a consequence of the epigenetic and metagenomic changes occurring in the gut, which are reflected in stool. Studies on human and mouse models have already demonstrated that long-term dietary changes drive stable modifications in the host epigenome and gut microbiota.42 43 In this respect, in our study, two stool DEmiRNAs (miR-636 and miR-4739) showed a clear downregulation with the duration of the non-omnivorous diet, independently of VT/VN subject’s age. MiR-636, an intragenic miRNA coded from the SKSF2 locus, has been found downregulated in blood/serum and adipose tissue of obese subjects44 and, on the contrary, upregulated in young subjects with obesity and diabetic nephropathy.45 46 In general, this miRNA seems to be implicated in weight loss. Garcia-Lacarte and colleagues,47 for example, found miR-636 hypomethylated and overexpressed in patients with metabolic syndrome who were highly responsive to the weight loss regimen. Presumably, the downregulation of this miRNA observed in VT and VN may be associated with a low-fat diet, ultimately more protective to the risk of obesity and diabetes.

MiR-4739 has been reported as a hub in the regulatory network mediated by the long non-coding RNA NEAT1.48 NEAT1 works as an miRNA sponge and represses the proliferation and fibrosis in diabetic nephropathy.49 Therefore, the action of NEAT1 as a natural downregulator of miR-4739 may be imitated by a VT or VN diet. Moreover, miR-4739 has been associated with the induction of adipogenic differentiation of human bone marrow stromal cells by targeting low-density lipoprotein receptor-related protein 3 (LRP3).50 Several genes related to metabolism, including SHMT1 and MKNK2, have been annotated as having the highest number of validated interactions with miR-636 and miR-4739. SHMT1 codes for serum hydroxymethyltransferase 1, a key protein involved in one-carbon metabolism.51 Conversely, MKNK2, encoding for MAPK interacting serine/threonine kinase 2, was recently associated with the lipid metabolism in adipocytes.52 Despite the fact that more data are needed to support an miRNA expression modulation on a long-term plant-based diet, our data highlight that in stool it is possible to detect miRNAs modulating metabolic processes, such as those of lipids, largely affected by different dietary regimes.

Some of the other stool DEmiRNAs found in this study are dysregulated in different diseases, as retrieved from HMDD, a database of experimentally supported human miRNA disease associations. MiR-143 was the DEmiRNA associated with the highest number of annotations (101 studies), followed by miR-124 (44 studies), miR-425 (11 studies) and miR-638 (10 studies). Interestingly, in our study, all these miRNAs were upregulated in stool of both VT/VN when compared with O. MiR-143 is an evolutionarily conserved miRNA transcribed as bicistronic RNA in a cluster with miR-145. MiR-143/miR-145 are well-known tumour suppressor miRNAs dysregulated in several cancer types, including CRC.53 Consistently, low levels of miR-143-3p have been observed in stool of patients with CRC54 55 and their upregulation in stool of VT/VN supports the lower cancer risk observed in relation to these dietary habits.56 57 These two miRNAs target some fundamental tight junction proteins.58 Nonetheless, miR-143-3p is a regulator of adipocyte differentiation59 and decreased levels of this miRNA were observed in obese individuals compared with normal weight controls in two independent studies.53 55 MiR-143/miR-145 cluster has an essential role in intestinal epithelium regeneration by modulating the insulin growth factor signalling pathway.60 Interestingly, in a mouse model of colitis, the administration of two probiotics, *Lactobacillus fermentum* and *L. salivarius*, ameliorated colonic damage and, concomitantly, increased miR-143 expression in colonic tissue.61 Conversely, miR-124 is active in colon tissues and associated with the modulation of an inflammatory response by regulating the signal transducer and activator of transcription 3 and acetylcholinesterase.62 63 MiR-124-5p is frequently downregulated in CRC tissues.64 In stool of VT/VN, increased expression levels of such miRNAs regulating the inflammatory response and the maintenance of gut epithelium are coherent with the known anti-inflammatory effects of plant-based diets.65

In the past decade, metagenomics has deeply increased our knowledge of the composition and function of the gut microbiome and how it is shaped by external factors.66 However, to the best of our knowledge, only a few studies in humans have explored the role of host miRNAs as mediators or interactors of the gut microbiome.67 In 2019, Virtue and colleagues68 have shown that miR-181 is overexpressed in white adipocytes of obese mice and knockout mice were less susceptible to a high-fat diet. In the same study, the authors showed that the expression of this miRNA is influenced by variations in the mouse gut microbiota. MiR-181 was also observed upregulated in the postprandial period of a high-saturated fat meal in peripheral blood mononuclear cells of healthy individuals68 supporting a link of this miRNA with lipid metabolism. Interestingly, in our study, miR-181c-3p stool levels progressively increased going from O to VN and were inversely correlated with the estimated animal lipid intake (Spearman ρ=-0.22). A differential expression of various lipid-related miRNAs in stool of VT/VN suggests that the change in dietary lipid intake may drive effective epigenetic changes in the gut, as previously proposed.69 In low-lipid dietary intervention studies, the stool levels of these miRNAs could represent a valid non-invasive parameter to assess the patient adherence and response to the diet.

In the present study, the gut microbiome taxonomic composition varied according to the dietary patterns, with *P. copri* and *Roseburia* sp. CAG 182, having a higher abundance in VT/VN than O. Interestingly, *A. muciniphila* abundance was linked to both diet and miRNAs, in agreement with recent observations.70 *A. muciniphila* is a well-known beneficial microbe involved in mucin metabolism whose levels are decreased in several diseases, including ulcerative colitis and metabolic disorders.71 Previous evidence reported that human miRNAs could directly target genes of these bacteria. Specifically, Liu and colleagues72 showed that the oral administration of human faecal miR-30c in a mouse model of multiple sclerosis ameliorated the disease symptoms by directly targeting *A. muciniphila* β-galactosidase coding-gene, which in turn increased the bacterial abundance in the gut. Despite miR-30c-5p was associated with low stool levels in our study population, it was significantly more abundant in stool of VT/VN.
Of note, higher stool levels of miR-425-3p, an miRNA upregulated in VT/VN, were significantly related to the abundance of A. muciniphila in our study. This miRNA was observed as significantly downregulated in serum of hyperlipidemic subjects. Furthermore, the other mature miRNA derived from miR-425 locus, miR-425-5p, was shown to be able to respond to a lipid challenge in mice small intestine and to inhibit 3-hydroxy-3-methylglutaryl-CoA synthase coding (HMGC32), a key enzyme of lipid metabolism. Finally, miR-425-5p overexpression in adipocytes was previously related to the regulation of lipogenesis and lipolysis. This data further strengthens our hypothesis on the role of the different dietary intake of lipids in VT/VN diets as a driver of consistent miRNA modulation in the gut. Higher stool levels of miR-425-3p and miR-638 were also significantly correlated with Roseburia and R. bacterium D5, which in our cohort were significantly more abundant in VT/VN, coherently with previous observation. This could be a further evidence of the important host-microbial interactions mediated by diet and epigenetic factors such as miRNAs. In this sense, our results on miRNAs could reflect the gut microbiota shifts in response to dietary fat recently reviewed by Chadaideh and Carmody.

In the present study, among the three investigated groups, the analysis of the dietary information recorded in the EPIC questionnaires confirmed the expected differences in weekly food consumption and estimated daily nutrient intake. Nutrients and specific diet components have been implicated in the modulation of miRNA expression in in vitro/in vivo models and, as a consequence, in all miRNA-mediated molecular mechanisms. In this study, vegetable lipids were the nutrients associated with the highest number of stool DEmiRNAs, together with animal lipids (namely miR-425-3p, miR-638, miR-8063 and miR-4277), but the latter showed an opposite trend. This result is consistent with previous evidence on the activity of lipids in modulating miRNA expression in different tissues, including gut, adipocytes, and plasma.

A functional enrichment analysis was performed to identify the candidate downstream signalling and metabolic pathways that may be affected by an intestinal miRNA expression modulation, which is reflected in stool samples. This analysis confirmed a relationship between the identified stool DEmiRNAs and nutrient metabolism. Terms related to fatty acid metabolism (namely, Glyoxylate and dicarboxylate metabolism and Phosphatidylglycerol metabolism) and those related to folate metabolism (eg, One carbon pool by folate, Metabolism of folate and pterines and Tetrahydrofolate metabolic process) were frequently observed among the targets of miRNAs upregulated in stool of VT/VN. In particular, the gene MTHFD2 coding for the mitochondrial bifunctional methylenetetrahydrofolate dehydrogenase/cyclohydrolase was predicted as a target of two stool miRNAs upregulated in both VT/VN (miR-425-3p, miR-4277) (online supplemental table 7). The estimated dietary intake of folic acid was also correlated with the stool levels of these miRNAs in our analysis. The activity of miRNAs in regulating folate metabolism has been previously observed by others. We can hypothesise that miR-425-3p and miR-4277 may form a negative feedback loop on the increasing level of this nutrient in VT/VN. Interestingly, miR-425-3p was included in the list of miRNAs regulated by the activity of nuclear factor erythroid 2-related factor 2 (Nrf2), a transcription factor involved in the regulation of metabolism downstream of nutrient-sensing pathways, whose activity is pivotal for the gastrointestinal tract development and maintenance.

MiRNAs are present in all eukaryotes: an intriguing aspect is that they could potentially ‘traverse’ from plants to animals through food via the gastrointestinal tract and access host cellular targets, where they could work as bioactive compounds and influence recipients’ physio-pathological conditions. However, this cross-kingdom interaction is still a topic of debate in the scientific community due to the lack of available data. Although some xenomiRs were detected in the present study, their low abundance in our data suggests a non-functional role of these molecules. Taking into consideration that xenomiR sequences may often be similar to those of human miRNAs and the absence of evidence on the capacity of xenomiRs to resist food processing thus impedes a definitive investigation.

This study is the first to integrate stool miRNAs with gut microbiota profiles and estimated daily nutrient intake from dietary questionnaires. Despite significant differences in BMI in the study population, this covariate only narrowly influenced our results. Most of the stool identified DEmiRNAs, including miR-425-3p, miR-638, miR-636, miR-143-3p, miR-181c-3p and miR-124-5p, remained significantly differentially expressed after adjustment for this variable. Similarly, although individuals have different dietary habits over the course of the year, correcting the analysis by time of sample collection only marginally affected the significances of the identified DEmiRNAs.

We also investigated miRNA profiles in plasma samples from the same subjects. However, in this biofluid, no significant DEmiRNAs were found according to the different dietary habits. It is possible that stool may reflect epigenetic changes occurring in the intestinal cells as a consequence of a stable dietary habit, while plasma samples could better reflect miRNAs characterising transient cellular response to nutrition-related stimuli, including circulating nutrients, hormones and microbial metabolites. This is consistent with previous observations in all the three dietary groups between the estimated intake of particular nutrients and a set of plasma miRNAs. A rising number of available data support the modification of faecal miRNA levels as a proxy of alteration and response occurring in the gut epithelium. Indeed, the main source of faecal miRNAs is represented by intestinal epithelial cells, as described by Liu et al. Although, in the present study, we did not assess the cell of origins of the detected miRNAs, the analysis of the TissueAtlas and Roadmap Epigenomics data confirmed that most of the stool DEmiRNAs identified in the present study are expressed and active in the gut. Interestingly, the analysis also showed a chromatin state related to a strong transcription at miR-143 coding locus coherently with previous observations. Then, we cannot exclude the contribution of other cellular components of the gastrointestinal systems, including those from the stromal and immune compartments. This is particularly relevant when trying to understand the host-microbiome crosstalk within the complex intestinal microenvironment. However, novel co-culturing systems and in vitro models reproducing the gut system could be applied based on evidence of miRNA and bacteria dysregulation observed in faecal samples to improve our understanding of the molecular characteristics of this miRNA-mediated host-microbial interaction.

In conclusion, we hereby present the first large-scale characterisation of faecal miRNAs in relation to different diets and to the gut microbial population with the state of the art of high-throughput technologies. We show a gradual modulation of the human miRNome as a consequence of different dietary regimens with important modifications in miRNAs involved in lipid and folate metabolism. Our data also provides novel evidence to improve our understanding of the role of small RNAs in the
host-microbial crosstalk in physiological and pathological conditions. An integrated analysis of faecal miRNA and gut microbiome signatures might provide insights for designing more accurate personalised dietary patterns and recommendations aimed at the prevention/treatment of chronic diseases. In particular, in monitoring the adherence to specific dietary regimes, where the intake of nutrients should be carefully checked, a translational approach of a non-invasive stool miRNA profiling may be foreseen.

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Acknowledgements
The authors are very grateful to all volunteers who participated with enthusiasm in the study. The authors are also thankful to Dr Vladimir Benes (Genecore Facility at EMBL, Heidelberg, Germany) for his kind collaboration and advice and to Mr Elton Jarvis Herman for his precious revision of the language of the text. The artwork for this study has been created with BioRender.com.

Contributors
BP, ST and AN contributed to concept and design; BP, GF, RGP, SG, ST, VP and AN to collection and assembly of the data; BP, GF, ST, AF, FC and AN to small RNA-Seq data analyses and interpretation; DE, FDF and EP to metagenomic data analyses and interpretation; ST, BP, GF, FC, DE, FDF and AN to manuscript writing; all authors gave final approval of the version.

Funding
This work was supported by Fondazione Umberto Veronesi postdoctoral fellowships 2017 and 2018 (recipients, ST and BP), Consorzio di Studio Interuniversitario ASSET (CSIA) grant number 2017-19 and by the JPI HDHL-ood, Diet, Intestinal Microbiomics and Human Health (ID 790) grant and the PRIN2017 (20174FHBWR_005) grant by the Italian Ministry of University and Research.

Interesting points
None declared.

Patient consent for publication
Not required.

Ethics approval
Azienda Ospedaliera "SS. Antonio e Biagio e C. Arrigo" of Alessandria. ID: colorectal_mirNA CEC2014.

Provenance and peer review
Not commissioned; externally peer reviewed.

Data availability statement
All data relevant to the study are included in the article or uploaded as supplementary information. Raw data are available upon request to the corresponding author.

Supplemental material
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