Supplementary Figure 1: a) m⁶A signals in the human and mouse XPO1 gene retrieved from the Metdb v2 database²⁹. Blue peaks correspond to meRIP-seq reads, yellow and green boxes are exons and UTR's respectively. Location of the CD-associated rs3087898 SNP is marked in the human transcript. b) Secondary structure of both forms of the human XPO1 5'UTR as predicted using the Vienna package³⁰. c) m⁶A RNA immunoprecipitation (meRIP) followed by RTqPCR of the 5'UTR of XPO1 and positive and negative controls in HCT116 intestinal and Jurkat T cell lines, n=3-5. (****p<0.0001; ns: non-significant according to a two-way ANOVA test. Enrichment relative to control IgG #p<0.05, ###p<0.001, ###p<0.0001 according to a two-way ANOVA test). d) Left, m⁶A levels in the 5'UTR of XPO1 in RNA extracted from human intestinal biopsies as assessed by meRIP-qPCR, n=3. (****p< 0.0001, according to a 2-tailed Student's ttest). Right, m⁶A RNA immunoprecipitation (meRIP) followed by RT-qPCR of the 5'UTR of XPO1 in immune (CD45+) and epithelial (CD326+) fractions from human intestinal biopsies, n=pool of 18 individuals. (**p<0.01 according to a two-way ANOVA test. Enrichment relative to control IgG #p<0.05, ns: non-significant according to a two-way ANOVA test). e) m⁶A-RT-qPCR of the 3 m⁶A motifs in the 5'UTR of XPO1 and positive and negative controls in intestinal cells, n=4. Significance was calculated relative to the negative control. (*p<0.05,**p<0.01 according to a oneway ANOVA test). f) Cellular localization of both XPO1 mRNA forms XPO1*C (C) and XPO1*T (T) and PO (cytoplasmic) and MALAT1 (nuclear) controls, n=4. (ns: non-significant according to a one-way ANOVA test). g) mRNA stability of both XPO1 mRNA forms XPO1*C (C) and XPO1*T (T) in cells treated with actinomycin D for 3 h and 6 h, n=3. (ns: non-significant according to a twoway ANOVA test). All values are means ± SEM.

Supplementary Figure 2: a) Relative expression of *METTL3* measured by RT-qPCR and **b)** overall m⁶A RNA levels measured by ELISA in cells transfected with an empty vector (EV) and a vector overexpressing METTL3 (ovMETTL3), n=3. (*p<0.05, ****p<0.0001 according to a two-tailed Student's t-test). **c-d)** Relative RNA expression measured by RT-qPCR and quantitative summary of the of *METTL3* and *YTHDF1* immunoblot in Figure 2B, (***p<0.001, ****p<0.0001 according to a two-tailed Student's t-test). **e)** overall m⁶A RNA levels measured by ELISA, (+p<0.1, ns:non-significant according to a two-tailed Student's t-test). **f)** m⁶A RNA immunoprecipitation (meRIP) followed by RT-qPCR of the 5'UTR of *XPO1* (*p<0.05; ns: non-significant according to a two-way ANOVA test. Enrichment relative to control IgG +p<0.1,

#p<0.05, ##p<0.01 according to a two-way ANOVA test) and **g)** quantitative summary of the XPO1 immunoblot in Figure 2C, n=3-4. (*p<0.05 according to a two-tailed Student's t-test). **h-i)** METTL3 and YTHDF1 immunoprecipitation RT-qPCR values for *XPO1* 5'UTR and m⁶A positive and negative controls, n=3-4. (Enrichment relative to control IgG +p<0.1, #p<0.05, ###p<0.001, ####p<0.0001, ns: non-significant according to a two-way ANOVA test). **j)** m⁶A immunoprecipitation (meRIP) RT-qPCR values for *XPO1* 5'UTR and m⁶A positive and negative controls in C26 mouse intestinal cells, n=3. (ns: non-significant according to a two-way ANOVA test. Enrichment relative to control IgG ###p<0.001 according to a two-way ANOVA test). **k)** Relative expression measured by RT-qPCR and quantitative summary of YTHDF1 immunoblot in Figure 2E and **I)** relative expression of *Xpo1* measured by RT-qPCR in WT (+/-) and *Ythdf1* knockout mice (DF1 KO), n=7. (**p<0.01, ***p<0.001, ns: non-significant according to a two-tailed Mann Whitney test). All values are means ± S.E.M.

Supplementary Figure 3: a) Schematic representation of the gliadin treatment set up in vitro (top) and in vivo (bottom). b) Left, overall m⁶A RNA levels measured by m⁶A Dotblot in cells, n=4. Right, quantitative summary of immunoblot in Figure 3A for XPO1, YTHDF1 and METTL3 in HCT116 cells, n=4. (+p<0.1, *p<0.05, **p<0.01, according to a two-tailed Student's t-test). c) Quantitative summary of XPO1 and YTHDF1 immunoblot in Figure 3C, n=3. (+p<0.1, ns:nonsignificant, according to a one-tailed Student's t-test). Silencing efficiency compared to NT cells (#p<0.05, according to a one-tailed Student's t-test). d) Left, overall m⁶A RNA levels measured by m⁶A Dotblot in WT control (NT) or PTG-treated mice (PTG), n=7. Right, quantitative summary of immunoblot in Figure 3D for XPO1 and YTHDF1 , n=7 (+p<0.1, *p<0.05, ns: non-significant according to a one-tailed Mann Whitney test). e) Left, overall m⁶A RNA levels measured by m⁶A Dotblot in humanized HLA-DR3-DQ2 control (NT) or PTG-treated mice (PTG). Right, quantitative summary of immunoblot in Figure 3D for XPO1 and YTHDF1, n=2-5 (+p<0.1, ns: non-significant according to a one-tailed Mann Whitney test). f) Quantitative summary of immunoblot in Figure 3F for YTHDF1 and XPO1 in untreated (-) and PTG-treated cells (+PTG) derived from wild type (WT) or Ythdf1 KO (KO) mice, n=2-4 (+p<0.1, ns:non-significant according to a one-way ANOVA test). All values are means ± S.E.M.

Supplementary Figure 4: a) Quantitative summary of immunoblot in Figure 4A for XPO1 and p50 in cells transfected with an empty vector (EV) and overexpressing both forms of XPO1 (C

and T), n=3. (*p<0.05, ***p<0.001 according to a two-way ANOVA test). b) Relative expression measured by RT-qPCR of XPO1 and IL8 in cells transfected with an empty vector (EV), overexpressing both forms of XPO1 (C and T) and cells overexpressing XPO1 and treated with the NFkB inhibitor Bay (T+BAY), n=3. (***p<0.001, ****p<0.0001, ns: non-significant according to a two-way ANOVA test). c) Representative immunoblot and quantitative summary of the immunoblot for XPO1 and p50 in untreated cells (NT) and cells treated with PTG, n=4. (*p<0.05, **p<0.01 according to a two-tailed Student's t-test). d) Quantitative summary of immunoblot in Figure 4F for YTHDF1, XPO1 and p50 in untreated cells (NT), cells treated with PTG and cells treated with PTG + silenced for YTHDF1 (PTG+siYTH), n=3. (+p<0.1, ns: non-significant according to a one-tailed Student's t-test). Silencing efficiency compared to NT cells (#p<0.05, according to a one-tailed Student's t-test), e) Quantitative summary of immunoblot in Figure 4G for XPO1 and p50 in untreated cells (NT), cells treated with PTG and cells treated with + XPO1 function inhibitor leptomycin B (PTG+LMB), n=3. (+p<0.1, *p<0.05, ns: non-significant according to a two-way ANOVA test). f) Relative expression measured by RT-qPCR of XPO1 and IL8 in untreated cells (NT), cells treated with PTG and cells treated with PTG and XPO1 function inhibitor leptomycin B (PTG+LMB), n=3. (*p<0.05, ****p<0.0001, ns: non-significant according to a twoway ANOVA test). Quantitative summary of immunoblot for p50 protein in g) WT and h) humanized HLA-DR3-DQ2 control mice (NT) or PTG-treated mice (PTG) in Figure 4J and K, respectively. n=7 and n=2-3 (*p<0.05 according to a one-tailed Mann Whitney test). i) Relative expression measured by RT-qPCR of mouse IL8 functional homolog cytokine Cxcl1 in untreated (-) and PTG-treated cells (+PTG) derived from wild type (WT) or Ythdf1 KO (KO) mice, n=2-4. All values are means ± S.E.M.

Supplementary Figure 5: a) Expression quantitative trait loci (eQTL) of *XPO1* expression levels compared between individuals with the protection genotype (CC) and individuals harboring the risk allele (CT+TT) n=7-25. Values are means ± S.E.M. (ns: non-significant, according to a two-tailed Mann Whitney test). **b)** RNA was extracted from untreated and PTG incubated biopsies and expression of *XPO1*, *METTL3* and *YTHDF1* was measured by RT-qPCR, n=15. Values are means of induction ± S.E.M. (+p<0.1, ns: non-significant according to a one-tailed Mann Whitney test). **c)** Correlation of the induction between *YTHDF1* and *XPO1* in PTG treated biopsies. Triangles correspond to *XPO1* mRNA and dots to XPO1 protein induction. n=9-15, R and p were calculated by Pearson correlation.

Supplementary Table1. List of used qPCR primers.

Gene	Primer Sequence	
XPO1 (5'UTR)	Fw: TGTTCCAGTCTTTGCTGCTG	
	Rv: AAGGCTCGCCTAAACTTTCC	
METTL3	Fw: TCGAGAGCGAAATTTTTCAAC	
	Rv: GGAGATAGAGAGCCTTCTGAACC	
YTHDF1	Fw: ACCTGTCCAGCTATTACCCG	
	Rv: TGGTGAGGTATGGAATCGGAG	
HPRT	Fw: ACCAGTCAACAGGGGACATAA	
	Rv: CTTCGTGGGGTCCTTTTCACC	
TUG1	Fw: ATTCCACGACCATGGTTGTC	
	Rv: ATTCACCACCACACACAG	
SOCS1	Fw: AGACCCCTTCTCACCTCTTG	
	Rv: AGTTAAGCTGCTACAACAACCAG	
IL8	Fw: ACTGAGAGTGATTGAGAGTGGAC	
	Rv: AACCCTCTGCACCCAGTTTTC	
5'UTR Xpo1	Fw: GGTGGGAAAACTGTGAAACCC	
	Rv: ACTGCTTCTTCCTTGTCC	
Ythdf1-/-	Fw: CCTGCATTCTCAGCATGG	
	Rv: GCTCCAGACTGTTCATCC	
Hprt	Fw: CTGGTGAAAAGGACCTCTCGAAG	
	Rv: CCAGTTTCACTAATGACACAAACG	
Mip2a	Fw: CCAACCACCAGGCTACAGG	
	Rv: GCGTCACACTCAAGCTCTG	
Cxcl1	Fw: CTGGGATTCACCTCAAGAACATC	
	Rv: CAGGGTCAAGGCAAGCCTC	

Supplementary Table 2. List of used qPCR assays.

Gene	Assay ID
XPO1	C16006954_10
MALAT1	Hs00273907_s1
RPLPO	Hs9999902_m1
Mettl3	Mm01316319_m1
Ythdf1	Mm00620538_m1
Rplpo	Mm00725448_s1

Supplementary Table 3. List of human biopsy samples.

ADULT INDIVIDUALS	Dx	ORIGIN	AGE (years)	SEX	HLA	Serology	Marsh (0-3)
	CD (n=6)	100% USA	34.2 ± 19.5	83.3% FEMALE		100% +	100% 3
				16.7% MALE			
	CTR	31.8% USA	45.2 ± 11.3	77.3% FEMALE	13.6% DQ2	100% -	100% 0
	(n=22)	68.2% SPAIN		22.7% MALE	4.5% DQ8		
	GFD		37.3 ± 11.4	33.3% FEMALE		100% -	83.3% 0
	(n=6)	100% USA		66.7% MALE			16.7% 1
PEDIATRIC INDIVIDUALS	Dx	ORIGIN	AGE (years)	SEX	HLA	Serology	Marsh (0-3)
	CD (n=28) 100% SPAIN	100% SPAIN	4.6 ± 3.7	48.5 FEMALE	69.7% DQ2	81.8% +	20% 0
		4.0 ± 3.7	51.5 % MALE	9.1% DQ2/DQ8	12.1% -	80% 3	
	CTR	CTR (n=16) 100% SPAIN	8.1± 3.6	68.75% FEMALE		100% -	100% 0
	(n=16)			31.25% MALE	-		
	GFD		69.2% FEMALE	69.2% DQ2	7.7% +	84.6% 0	
	(n=12)	100% SPAIN	2 ± 0.7	30.8% MALE	23.1% DQ2/DQ8	92.3% -	7.7% 1 7.7% 3